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Novel morpholin-3-one fused quinazoline derivatives as EGFR tyrosine kinase inhibitors

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EGFR^{wt}: IC₅₀ 53.1 nM
EGFR^{T790M/L858R}: IC₅₀ 0.336 μ M
H358 cell: IC₅₀ 13.46 μ M
A549 cell: IC₅₀ 7.51 μ M

Novel morpholin-3-one fused quinazoline derivatives as EGFR tyrosine kinase inhibitors

Xuemei Qin^a, Yongjuan Lv^a, Peng Liu^b, Zhipeng Li^a, Liming Hu^{a,*}, Chengchu Zeng^a, and Leifu Yang^c

^a College of Life Science and Bioengineering, Beijing University of Technology, Beijing, 100124, China

^b Guangzhou Institute of Biomedicine and Health, Chinese Academy of Science, Guangzhou, 510530, China

^c Beijing Dalitai Pharmaceutical Technology Co., Ltd, Beijing, 100176, China

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ABSTRACT

A series of novel morpholin-3-one-fused quinazoline derivatives were designed, synthesized and evaluated as EGFR tyrosine kinase inhibitors. Nineteen compounds showed significant inhibitory activities against EGFR^{wt} kinase ($IC_{50} < 1 \mu M$). Compound **a8** demonstrated the most potent inhibitory activity toward EGFR^{wt} ($IC_{50} = 53.1 \text{ nM}$). Compound **a7** and **a8** showed excellent inhibitory activities against mutant EGFR^{T790M/L858R} and strong antiproliferative activity against H358 and A549 cell lines. Finally, molecular docking studies were performed to predict the possible binding mode of the target compounds. It is believed that this work would be very useful for designing a new series of tyrosine kinase inhibitors targeting EGFR.

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The epidermal growth factor receptor (EGFR) tyrosine kinase is a key mediator in cellular signalling transduction pathways and is closely related to cell proliferation, survival, adhesion, migration, and differentiation.^{1,2} Deregulation of EGFR signalling has been observed in many human cancers, including colorectal, head and neck, lung, breast, ovarian, and bladder cancers.³ Thus, EGFR and its family members have emerged as attractive targets for anti-cancer therapy, especially for the treatment of non-small-cell lung cancer (NSCLC).^{4,5} Several EGFR inhibitors, such as gefitinib and erlotinib (Fig. 1) belonging to a class of 4-anilinoquinazolines have made significant progress in the treatment of NSCLC patients harboring somatic EGFR mutations L858R and delE746_A750.⁶⁻⁸ However, the undeniable success of EGFR tyrosine kinase inhibitors are limited by the up-regulation of bypass signalling pathways and acquired point mutations.^{9,10} After treatment 10-16 months with these inhibitors in western countries, approximately 50% of the NSCLC patients gain an additional gate keeper mutation (T790M).^{11,12} The secondary T790M mutation decreases sensitivity to gefitinib or erlotinib and increases the binding affinity for ATP.^{13,14}

In order to overcome the T790M mutation related resistance, several irreversible EGFR second-generation covalent inhibitors (Fig. 1), which form a covalent bond with Cys797 within the EGFR active site, were developed.¹⁵⁻¹⁷ Afatinib was the first approved irreversible EGFR inhibitor by the US FDA for the treatment of late-stage NSCLC patients with actively mutated

EGFR.¹⁸ However, these irreversible EGFR inhibitors lacked significant selectivity between EGFR T790M (EGFR^{T790M}) mutants and the wild-type kinase that leads to dose-limiting toxicities in these patients, including skin rash and diarrhea that can result in severe dehydration and kidney failure.^{14,19} Recently, third-generation covalent inhibitors including AZD9291 and CO-1686 have been generated that demonstrate selectivity for EGFR^{T790M} mutants over wild-type EGFR (EGFR^{wt}), and early phase I data indicated promising efficacy.²⁰⁻²² Although, the number of potent EGFR inhibitors is constantly expanding, discovery of inhibitors with novel scaffolds remains a focus nowadays. Herein, we would like to report the design, synthesis and biological evaluation of morpholin-3-one fused quinazoline derivatives as new EGFR inhibitors.

The co-crystal structures of gefitinib or erlotinib^{8,23} with EGFR have provided valuable structural information for helping the rational design of potent EGFR inhibitors. Binding mode analysis of gefitinib suggests several key interactions with the ATP binding pocket of EGFR, including the critical H-bond of quinazoline moiety with the hinge region, the hydrophobic pocket filled up by the 3-chloro-4-fluoro aniline substituent and the solvent region occupation of 6-morpholinopropoxy group. Solvent region generally accommodate a variety of polar functional groups with little affection on the efficacy.²⁴ The success of icotinib (Fig. 1), which mimics erlotinib with a crown ether fused quinazoline, is a good example of this strategy.^{25,26}

* Corresponding author. Tel.: +86-10-6739-6211; fax: +86-10-6739-6211; e-mail: huliming@bjut.edu.cn

Accordingly, we have designed and synthesized a series of novel morpholin-3-one fused quinazoline derivatives through intramolecular cyclization as new EGFR inhibitors (Fig. 2).

The synthetic route of compounds **a1-a15** is illustrated in Scheme 1. Commercially available 4-hydroxy-3-nitrobenzoic acid **1** was esterified with methanol using concentrated sulfuric acid as a catalyst to give compound **2**.²⁷ Alkylation of methyl 4-hydroxy-3-nitrobenzoate **2** with ethyl bromoacetate and reduction of the nitro group with Fe/acetic acid followed by in situ cyclization yielded intermediate **4**. Subsequent nitration and alkylation provided **6**. Then reduction the nitro group of **6** using Fe/acetic acid gave **7**, which was subjected to cyclization to generate intermediate **8**. Following, **8** was chloridized by phosphorus oxychloride to generate intermediate **9**. Finally, condensation of **9** with substituted anilines afforded 4-anilinoquinazoline derivatives **a1-a15**.

Synthesis of compounds **b1-b6** is shown in Scheme 2. Intermediate **5**, which could be synthesized according to scheme 1, was reacted with 1-bromo-3-chloropropane and piperidine in

sequence to achieve **11**. The target compounds were synthesized in good yields by the next four step reactions according to scheme 1.

To evaluate the EGFR inhibitory potency of the new compounds, Kinase-Glo luminescent assay is used to test their ability to block EGFR tyrosine kinase. Compounds were initially tested at final concentrations of 1000, 200, 40, 8, 1.6, 0.32, and 0.06 nM and performed in duplicate at each concentration with gefitinib as positive control. Compound inhibition curve was

Fig. 2. Design strategy and modification of novel EGFR inhibitors.

Fig. 1. Chemical structures of EGFR-TKIs.

Scheme 1. Synthetic route of the target compounds **a1-a15**. Reagents and conditions: (a) CH₃OH, conc. H₂SO₄, reflux; (b) ethyl bromoacetate, K₂CO₃, DMF, 70 °C; (c) Fe, HAc, 70 °C; (d) HAc/HNO₃; (e) 4-(3-chloropropyl)morpholine, K₂CO₃, KI, DMF, 80 °C; (f) Fe, HAc, EtOH/H₂O; (g) formamidine acetate, EtOH, reflux; (h) POCl₃, 120 °C; (i) anilines, isopropanol, reflux.

Scheme 2. Synthetic route of the target compounds **b1-b6**. Reagents and conditions: (a) 1-bromo-3-chloropropane, Cs_2CO_3 , acetonitrile, rt; (b) piperidine, K_2CO_3 , KI, DMF, rt; (c) Fe, HAc, EtOH/ H_2O ; (d) formamidine acetate, EtOH, reflux; (e) POCl_3 , 120 °C; (f) anilines, isopropanol, reflux.

fitted using Graphpad Prism 5.0 software based on the inhibition rate of all compounds at different concentrations. The testing results are summarized in Table 1. Almost all the compounds displayed significant EGFR inhibitory activities ($\text{IC}_{50} < 1 \mu\text{M}$). Among them, compound **a8** showed the most potent inhibitory activity against EGFR^{wt} kinase ($\text{IC}_{50} = 53.1 \text{ nM}$). When R was 3-morpholinopropyl (**a1-a15**), the inhibitory activities against EGFR^{wt} ranged from 53.1 nM to 829.2 nM except **a5** and **a14** in which $\text{IC}_{50} > 1000 \text{ nM}$. When R¹ was halogen (F, Cl or Br) and R² was hydrogen (**a1**, **a2**, **a3**), compounds substituted by Cl or Br (**a2**, $\text{IC}_{50} = 63.1 \text{ nM}$; **a3**, $\text{IC}_{50} = 89.6 \text{ nM}$) displayed 2-fold increase in potency than substituted by F (**a1**, $\text{IC}_{50} = 245.1 \text{ nM}$). While R¹ was hydrogen and R² was halogen atom (**a9**, **a10**, **a11**), the results of inhibition against EGFR was unsatisfactory. This may be that meta-position compounds (**a1**, **a2**, **a3**) could form a stronger interaction with EGFR kinase than para-position compounds (**a9**, **a10**, **a11**). When R¹ was Cl and R² was F, compound **a8** displayed the most inhibitory activity toward EGFR^{wt} ($\text{IC}_{50} = 53.1 \text{ nM}$). When R¹ or R² was methoxy or trifluoromethyl group, compounds (**a5**, **a6**, **a14**), displayed weaker potency against EGFR kinase. One explanation might be that a larger volume of R¹ group could not effectively fill up well the hydrophobic pocket of EGFR kinase. In summary, when R¹ was Cl or Br, and R² was H or F, compounds (**a2**, **a3**, **a8**) displayed excellent inhibitory activities against EGFR^{wt} ($\text{IC}_{50} < 89.6 \text{ nM}$). Among them, compound **a8** displayed the most inhibitory activity toward EGFR^{wt} ($\text{IC}_{50} = 53.1 \text{ nM}$).

To find a more potent EGFR inhibitor, R was substituted by 3-(piperidin-1-yl)propyl in the subsequent synthesis (**b1-b6**). Among them, **b3** was the most active compound against EGFR ($\text{IC}_{50} = 99 \text{ nM}$). Compounds **b2**, **b3**, **b6** (R¹ = Cl or Br) showed much more potency than **b1** (R¹ = F). This was the same phenomena as the former discussion (**a1-a3**, R¹ = halogen). The reason may be that small fluorine atom could be too small to fill up well the hydrophobic pocket of EGFR kinase pocket. However, when R¹ was a relatively larger volume group methyl or ethynyl, compounds (**b4**, **b5**) did not showed good inhibitory activities towards EGFR^{wt}. As a whole, R¹ was Cl or Br would be beneficial for improving the inhibitory activities against EGFR^{wt}.

By comparing **a1&b1**, **a2&b2**, **a3&b3**, **a4&b4**, **a7&b5** and **a8&b6**, it is easy to find that compounds with R being 3-morpholinopropyl show much more inhibitory efficiency against EGFR^{wt} than compounds with R being 3-(piperidin-1-yl)propyl do. This indicates that the increased hydrophily of 6-position substituent of quinazoline may be beneficial for enhancing the inhibitory activity against EGFR.

Among all the compounds, **a7** and **a8** were picked up for further biological evaluation. One reason was their good inhibitory activities against wip-type EGFR. The other reason was that 4-position substituent of quinazoline was 3-chloro-4-

Table 1. In vitro enzymatic inhibitory activities of compounds **a1-a15** and **b1-b6** against EGFR^{wt}

		a		b	
Compound	R ¹	R ²	EGFR ^{wt} IC ₅₀ (nM)		
a1	F	H	245.1		
a2	Cl	H	63.1		
a3	Br	H	89.6		
a4	Me	H	186.9		
a5	MeO-	H	>1000		
a6	-CF ₃	H	829.2		
a7	ethynyl	H	139.0		
a8	Cl	F	53.1		
a9	H	F	434.1		
a10	H	Cl	353.7		
a11	H	Br	823.9		
a12	H	Me	594.5		
a13	H	H	518.6		
a14	MeO-	MeO-	>1000		
a15	Cl	Me	162.7		
b1	F	H	460.0		
b2	Cl	H	115.9		
b3	Br	H	99.0		
b4	Me	H	456.2		
b5	ethynyl	H	211.4		
b6	Cl	F	137.9		
gefitinib			11.8		

Table 2. Inhibition of EGFR^{T790M/L858R} kinase, H358 and A549 Cell Lines by **a7**, **a8** in vitro

Compound	EGFR ^{T790M/L858R} IC ₅₀ (μM)	H358 IC ₅₀ (μM)	A549 IC ₅₀ (μM)
a7	0.880	13.49	5.58
a8	0.336	13.46	7.51
gefitinib	0.025	14.66	9.28
erlotinib	0.068	6.85	6.26

fluoro or 3- ethynyl aniline, which was found in many marketed drugs as an important group and were studied widely in different areas. The third reason was that target compounds showed much more inhibitory efficiency to EGFR^{wt} when R was 3-morpholino-propyl than when R was 3-(piperidin-1-yl)propyl.

In the T790M/L858R mutant EGFR kinase assays (Table 2), compound **a8** (IC₅₀ = 0.88 μM) was about 2-fold more potent than **a7** (IC₅₀ = 0.336 μM). The two compounds showed significant inhibitory activities against EGFR^{T790M/L858R} compared to the standards erlotinib and gefitinib.

To experiment the antiproliferative activity of **a7** and **a8**, both compounds were tested on H358 (human NSCLC cell line) and A549 (a human lung adenocarcinoma cell line). As showed in Table 2, **a7** and **a8** strongly suppressed the proliferation of H358 and A549 cells compared to gefitinib and erlotinib.

To gain more understanding of the structure-activity relationships observed for EGFR kinase, molecular docking of the **a8** into ATP binding site of EGFR kinase was performed on the binding model based on the EGFR/gefitinib complex crystal structure (2ITY.pdb). Docking mode appeared superimposition of the **a8** with the control ligand gefitinib in a similar binding mode. The morpholin-3-one fused scaffold was located well in the ATP pocket. One conventional hydrogen bond was observed between the nitrogen of the quinazoline ring and the hydrogen of Met793 amide and another water mediated hydrogen bond was observed between oxygen on the quinazoline ring and the amide nitrogen of Thr791. This may be the reason that **a8** showed wonderful inhibitory activity against EGFR kinase. And we can observe from Fig. 3 the morphin-3-one group of **a8** is shifted away from the morphine of gefitinib. However, it is believed that the charge-charge interaction between basic nitrogen in morpholin-3-one group of gefitinib and Asp800 is very important for gefitinib binding into EGFR active site. This may be the reason why **a8** is less potent activities to EGFR kinase than gefitinib in its inhibitory activity against EGFR kinase. Therefore, one way to improve the inhibitory activities for these

Fig. 3. Docking modes of compound **a8** (gray) overlaid with the binding conformation of gefitinib (yellow) into the EGFR kinase domain compounds could be to link the amine group with different linkers.

In summary, a series of novel morpholin-3-one fused quinazoline derivatives were designed, synthesized and their biological activities were evaluated. Most compounds showed good inhibitory activities against EGFR^{wt} kinase (IC₅₀ < 1 μM). Among them, compound **a8** demonstrated the most potent inhibitory activity (IC₅₀ = 53.1 nM against EGFR^{wt}). Compounds **a7** and **a8** showed good inhibitory activities against mutant EGFR^{T790M/L858R} and strong antiproliferative activity against H358 and A549 cell line compared to gefitinib and erlotinib. Docking modes of compound **a8** overlaid well with the binding conformation of gefitinib in the EGFR kinase domain. It is believed that this work would have important implications for the design of new tyrosine kinase inhibitors targeting EGFR.

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Supplementary Material

Supplementary material that may be helpful in the review process should be prepared and provided as a separate electronic file. That file can then be transformed into PDF format and submitted along with the manuscript and graphic files to the appropriate editorial office.

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