Synthesis and biological evaluation of novel 2,3-dihydrochromeno[3,4-d]imidazol-4(1H)-one derivatives as potent anticancer cell proliferation and migration agents

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ABSTRACT: In this study, a series of novel molecules containing chromeno [3,4-d]imidazol-4(1H)-one was synthesized and their biological activities were evaluated. Among them, compound **35** showed a dramatic anticancer activity against HCT116 and MCF-7, and the flow cytometry assays demonstrated that it could arrest G0/G1 cell-cycle and induce apoptosis of SW620 cells in a dose-dependent manner. Besides, it also blocked MCF-7 cancer cell migration. Moreover, it inhibited tumor growth in HCT116 subcutaneously implanted xenografted mice. Taken together, compound **35** may be a promising candidate for anti-cancer agent as well as metastatic one.

Keywords: chromeno [3,4-d]imidazol-4(1H)-one; anti-cancer agents, anti-metastasis

1. Introduction

Cancer is becoming a leading cause of death in developing and developed countries. An estimated 14.1 million new cancer cases occurred in 2012 worldwide and Eastern Asia accounted for 29.4% of it [1]. There were 8.2 million cancer deaths in 2012 all over the world, and cancer has also been the first reason of death in China under the age of 65 [1, 2].

A pharmacophore hybrid approach for exploration of novel and highly active compounds is an effective and commonly used direction in modern medicinal chemistry. Hybridization of two different bioactive molecules with complementary pharmacophoric functions often showed synergistic effects [3, 4]. For instance, pyrazole [5] and stilbene [6] conjugated with coumarin derivatives possess good antiproliferative activities. Given the important role of this fusion strategy, previously, we also synthesized a series of triazol and coumarin fused derivatives which showed efficient anticancer activities [7].

The coumarin and its derivatives are widely used for antitumor agents [8-11]. The structural modification and structure-activity relationship analysis of coumarins on anticancer effect have

been fueled by various academics and industries [12-14]. Another prevalent scaffold, imidazole template, is a privileged structure fragment in current medicinal chemistry [15-21]. In particular, benzimidazole and imidazoquinoline derivatives are well known for their pharmaceutical properties, which have been extended to widespread inhibitors of proton pump [22], hypertension, virus [23] and cancer [24-26]. (Fig. 1.) Because there are few reports on the biological research on the coumarin and imidazole fused skeleton, in this study, a series of compounds possessing a chromeno [3,4-d] imidazol-4(1*H*)-one core was synthesized and their structure-activity relationships (SAR) were studied.

2. Results and discussion

2.1 Structure and activity relationship

At the beginning of our research, the coumarin analogues were substituted by the simple phenyl on the N-1 position and aryl fragments on C-6 position. As displayed in Table 1, phenyl groups including pyridin-3-yl, 3-NH₂-C₆H₄, 2-Me-C₆H₄, 4-OH-C₆H₄ and 3-NHSO₂-(4-F-C₆H₄)-C₆H₄ on the C-6 position showed no bioactivity. When the pyridin-4-yl and quinolin-3-yl substituted the C-6 position of coumarin, the inhibitory activities of compounds **15** and **16** against three tested cancer cell lines were improved.

Furthermore, we turned our attention to the phenyl ring at the N-1 position. Compounds **17** - **25** with 4-CH₂CN in the phenyl ring displayed slight improvement in inhibitory activities against U87-MG, PC-3 and HCT116 cancer cell lines. Specifically, when 3-NH₂-C₆H₄ was introduced to the C-6 position, compound **19** showed moderate activity against PC-3 cell line with IC₅₀ value of 36.51 μM. Furthermore, compound **21** that containing a 4-OH-C₆H₄, exhibited some bioactivity with value of 29.37 μM. Besides, pyridine-3-yl (**22**) or quinolin-3-yl (**25**) at C-6 position showed better activities with IC₅₀ values of 29.37 μM and 28.4 μM against HCT116 cells, indicating that the nitrogen atom on the heterocyclic substitute is crucial to the bioactivity, which may provide a hydrogen acceptor site.

To further investigate the SAR of N-1 group, with pyridin-4-yl, pyridine-3-yl or quinolin-3-yl retained at C6-position, the phenyl was replaced by 4-OMe-C₆H₄ and 2-Me-C₆H₄. However, all the compounds (compounds **26** to **31**) showed no bioactivity, suggesting that the cyano of 4-CH₂CN-C₆H₄ group is a very crucial moiety which may provide a hydrogen acceptor site. When N4-C(CH₃)₂-C₆H₄ group was introduced to the N-1 position, all compounds showed an improved inhibitory activities. Compound **32** with pyridin-3-yl at C-6 position showed antitumor activity against U87-MG cells with IC₅₀ value of 25.62 μM and **33** inhibited U87-MG and PC-3 with IC₅₀ values of 28.55 μM and 29.01 μM, respectively. What's more, **34** with quinolin-3-yl displayed higher bioactivity against the U87-MG, PC-3 and HCT116 cell lines with IC₅₀ values of 33.42, 19.76, 20.40 μM.

Out of our expectation, when a methyl group was attempted to introduce to N-3 position, tert-butoxy was accidentally inserted into C-2 of imidazole ring. To our delight, compound **35** possesses attractive antitumor activity against HCT116 with IC₅₀ value of 1.40 μM, 15 fold stronger than that of **34**. (Table 2) It can be concluded that the good inhibitory activity does not simply reside in the quinolin-3-yl at C-6 position, but also the methyl and tert-butoxy at the ring of imidazole.

2.2 Biological evaluation

2.2.1 Compound **35** inhibited many kinds of cancer cells with low toxicity to normal LO2 cells

In order to investigate whether **35** has widely anti-cancer activities, the proliferation inhibition on colon carcinoma SW420 cell, CT26 cell and HCT116 cell, human breast carcinoma MCF-7 cell and lung carcinoma A549 cell were tested. As a result, compound **35** displayed attractive antitumor activities against SW620, CT26, HCT116, MCF-7, and A549 with IC₅₀ values of 3.55, 3.50, 1.40, 1.70 and 9.50 μ M, respectively (Fig. 2A). Meanwhile, it also expressed relatively low cytotoxicity to LO2 normal cell with the IC₅₀ value of 37.0 μ M.

2.2.2 Compound **35** inhibited the formation of colony in HCT116

As indicated in Fig. 2B, after the treatment of **35**, the size of the colonies was apparently smaller than that of the non-treated group. When the concentration of **35** was increased to 2.50 μ M, nearly no colony formation was observed.

2.2.3 Compound **35** induced G0/G1 phase arrest and apoptosis

The effects of **35** on SW620 cell cycle distribution were shown in Fig. 3A. Under the treatment of **35**, SW620 cells were arrested significantly at the G0/G1 phase compared to the untreated group, with the percentage of G0/G1 increased from 38.47% (control) to 38.78% (0.3125 μ M), 40.30% (0.625 μ M), 42.97% (1.25 μ M), 47.49% (2.50 μ M) and 60.04% (5.0 μ M), respectively. Meanwhile, the number of cells in S and G2/M phase decreased. Furthermore, flow cytometry analysis using Annexin V-FITC and propidium iodide (PI) staining showed that **35** treatment at 0.3125, 0.625, 1.25, 2.50 and 5.0 μ M increased the percentage of apoptosis cells from 2.83% to 15.71%, 25.19%, 26.22% and 42.90%, respectively. From the above experiments, it can be concluded that **35** could induce G0/G1 arrest and apoptosis of SW620 cells in a dose-dependent manner.

2.2.4 Compound **35** blocked MCF-7 cancer cell migration

To further test the effect of **35** on the cancer cell migration potential, wound healing assay was conducted on MCF-7 cells. As shown in Fig. 4, after treatment with **35** at 5.0 μ M for 24 h, the breast cancer cell migration was suppressed obviously.

2.2.5 Compound **35** inhibited tumor growth in HCT116 mouse mammary tumor model

To assess the antitumor effect of **35** *in vivo*, nude mice were administered **35** by subcutaneous injection at the dose of 40 mg/kg and 80 mg/kg. As shown in Fig. 5A, **35** markedly suppressed tumor growth in a dose-dependent manner with the inhibition of tumor progression at 52.96% in the 80 mg/kg group. In addition, **35** treatment was well tolerated and did not cause significant body weight loss (Fig. 5B).

3. Conclusion

In this research, a series of chromeno [3,4-d] imidazol-4(1*H*)-one derivatives was synthesized and their antitumor activities *in vitro* against U87-MG cells, PC-3 cells and HCT116 cells were

tested. Among these compounds, **35** exhibits excellent broad spectrum anticancer activities *in vitro* with IC₅₀ values of 1.40 μM, 3.55 μM, 1.70 μM, 3.50 μM and 9.50 μM against HCT116, SW620, MCF-7, CT26 and A549 respectively. In addition, structure activity relationship research reveals that 4-C(CH₃)₂-CN-C₆H₄ at N-1 position of coumarin core is the best optimal for bioactivity; a methyl group at N-3 position and tert-butoxy inserted into C-2 of imidazole are indispensable for the improvement of potency; the quinolin-3-yl at C-6 position of coumarin core can make a positive contribution to the activity. Flow cytometry experiment demonstrates that compound **35** can arrest the cell cycle of SW620 at G0/G1 and induce apoptosis in a concentration-dependent manner. Further *in vivo* anticancer experiments on subcutaneous HCT116 tumor model in nude mice show that **35** can inhibit tumor growth by 52.96% at 80 mg/kg/48h for 20 days. Altogether, our results provide a novel scaffold with better potency as antitumor agents. All the results encourage us to continue the development and the further validation of mechanism at the molecular level.

4. Experimental section

4.1 General methods for chemistry

All the reagents and solvents were purchased from commercial vendors and dried by standard methods in advance and distilled before used. ¹H and ¹³C NMR spectra were recorded on with a Bruker AC-E400 spectrometer (operating at 400 MHz), with chemical shift in parts per million (ppm, δ) downfield from TMS as an internal standard. Mass spectrometry (MS) data were obtained using a BrukerAmjzon spectrometer. Compound purity was determined by a Waters 1525 series HPLC system with a confirming purity ≥95% for all of the final biologically tested compounds. Flash column chromatography was completed by silica gel.

3-(4,5)-dimethylthiahiazol(-z-yl)-3,5-di-phenyltetrazolium bromide (MTT), DMSO, DMEM, were from Sigma. The Annexin V-FITC Apoptosis Detection Kit was purchased from KeyGEN BioTECH.

4.2 General procedure for the synthesis of **1** – **5**

4.2.1 Synthesis of **1**

The mixture of 4-bromophenol (116 mmol) and acetic anhydride (464 mmol) in pyridine (60 mL) was stirred at 100 °C for 3.5 h. Then the reaction mixture was cooled to room temperature and quenched with water. The reaction was acidized with hydrochloric acid and extracted with ethyl acetate (40 mL×3). The combined organic layer was washed by saturated sodium chloride solution for three times, dried over anhydrous Na₂SO₄ and concentrated under reduced pressure. The residue was purified by silica gel chromatography. **1**: yield 100%.

4.2.2 Synthesis of **2**

A suspension of **1** (46 mmol) and aluminum chloride (69 mmol) was stirred at 150 °C. After 3.5 h the mixture was cooled to room temperature and the diluted hydrochloric acid was added to it slowly for quenching aluminum chloride until all solid disappeared. Then the solution was extracted with ethyl acetate (20 mL×3), and the combined organic layer was washed by saturated

sodium chloride solution for three times, dried over anhydrous Na_2SO_4 and concentrated under reduced pressure. The decolorized solid was recrystallized with n-hexane. **2**: yield 72%. ^1H NMR (400 MHz, CDCl_3): δ 7.38 (d, J = 8.0 Hz, 1H), 7.32 (s, 1H), 7.23 (d, J = 7.9 Hz, 1H), 7.06 (d, J = 8.2 Hz, 1H), 2.28 (s, 3H) ppm.

4.2.3 Synthesis of **3**

2 (74 mmol) and diethyl carbonate (111 mmol) in toluene (160 mL) were added dropwise to sodium hydride (369 mmol) in toluene (50 mL) in ice-water bath. The mixture was stirred at ice bath for 30 min and at 100 °C for 4 h. The sodium hydride was quenched with water, acidized with hydrochloric acid and then the suspension was filtered separately, and the solid collected from this filtration was washed with toluene and water for three times. The solid dried at room temperature with phosphorus pentoxide. **3**: yield 71%. ^1H NMR (400 MHz, CDCl_3): δ 12.17 (s, 1H), 7.83 (d, J = 2.45, 1H), 7.55 (dd, J = 2.4 Hz, 8.8 Hz, 1H), 6.90 (d, J = 9.2 Hz, 1H), 2.63 (s, 3H) ppm.

4.2.4 Synthesis of **4**

6-bromo-4-hydroxy-2H-chromen-2-one (0.15 mol) was dissolved in dichloromethane (300 mL) and then stirred at room temperature. The solution was acidified with hydrochloric acid and the yellow solid was filtered off and subsequently washed with water and acetic acid. **4**: yield 94%. ESI-MS: Calcd for $\text{C}_9\text{H}_4\text{BrNO}_5$ $[\text{M}-\text{H}]^-$: 463.98, Found 284.93, 286.93, found 283.9, 285.9. ^1H NMR (400 MHz, $\text{d}_6\text{-DMSO}$): δ 7.93 (d, J = 2.4 Hz, 1H), 7.68 (dd, J = 2.4 Hz, 8.8 Hz, 1H), 7.84 (d, J = 8.8 Hz, 1H) ppm.

4.2.5 Synthesis of **5**

To a suspension of 6-bromo-4-hydroxy-3-nitro-2H-chromen-2-one (10 mmol) in phosphorus oxychloride (15 mL) was added triethylamine slowly. The mixture was stirred and refluxed for 1 h. The phosphorus oxychloride was quenched with water carefully, then the suspension was filtered and the decolorized solid was recrystallized with ethyl acetate and petroleum ether. **5**: yield 87%. ^1H NMR (400 MHz, $\text{d}_6\text{-DMSO}$): δ 7.92 (d, J = 2.4 Hz, 1H), 7.67 (dd, J = 2.6 Hz, 9.0 Hz, 1H), 7.18 (d, J = 8.8 Hz, 1H) ppm.

4.3 General procedure for the synthesis of **6a** - **6e**

A mixture of corresponding phenylamine (20 mmol) and **5** (20 mmol) was added into DMF (50 mL). The reaction mixture was stirred at room temperature for 5 h. Then, it was diluted with water and the separated solid was filtered off, washed with water, ethanol and ethyl acetate.

4.3.1 6-bromo-3-nitro-4-(phenylamino)-2H-chromen-2-one (**6a**)

Yield: 95%. ESI-MS: Calcd for $\text{C}_{15}\text{H}_9\text{BrN}_2\text{O}_4$ $[\text{M}-\text{H}]^-$: 358.97, 360.98, Found 358.9, 360.9. ^1H NMR (400 MHz, DMSO) δ 10.36 (s, 1H), 8.35 (d, J = 8.7 Hz, 1H), 7.84 (d, J = 1.7 Hz, 1H), 7.74 (dd, J = 8.7, 1.7 Hz, 1H), 7.37 (t, J = 7.7 Hz, 2H), 7.25 (t, J = 7.4 Hz, 1H), 7.20 (d, J = 7.7 Hz, 2H) ppm.

215
216 4.3.2 2-(4-((6-bromo-3-nitro-2-oxo-2H-chromen-4-yl)amino)phenyl)acetonitrile (**6b**)

217 Yield: 93%. ESI-MS: Calcd for C₁₇H₁₀BrN₃O₄ [M+Na]⁺: 421.99, 423.99, Found 422.0, 423.7.
218 ¹H NMR (400 MHz, DMSO): δ 10.37 (s, 1H), 8.74 (s, 1H), 7.96 (d, *J* = 7.2 Hz, 1H), 7.47 (d, *J* =
219 8.8 Hz, 1H), 7.35 (d, *J* = 8.4 Hz, 2H), 7.23 (d, *J* = 8.4 Hz, 2H), 4.08 (s, 2H) ppm.

220
221 4.3.3 6-bromo-4-((4-methoxyphenyl)amino)-3-nitro-2H-chromen-2-one (**6c**)

222 Yield: 95%. ESI-MS: Calcd for C₁₆H₁₁BrN₂O₅ [M-H]⁻: 388.99, 390.98, Found 389.0, 391.1. ¹H
223 NMR (400 MHz, DMSO): δ 10.25 (s, 1H), 8.33 (d, *J* = 8.8 Hz, 1H), 7.82 (d, *J* = 1.8 Hz, 1H), 7.73
224 (dd, *J* = 8.7, 1.8 Hz, 1H), 7.15 (d, *J* = 8.9 Hz, 2H), 6.92 (d, *J* = 8.9 Hz, 2H), 3.77 (s, 3H) ppm.

225
226 4.3.4 6-bromo-3-nitro-4-(*o*-tolylamino)-2H-chromen-2-one (**6d**)

227 Yield: 97%. ESI-MS: Calcd for C₁₆H₁₁BrN₂O₄ [M+Na]⁺: 396.99, 398.99, Found 397.2, 399.2.
228 ¹H NMR (400 MHz, CDCl₃) δ 11.43 (s, 1H), 7.63 (d, *J* = 8.8 Hz, 1H), 7.42 (m, 2H), 7.32 (t, *J* =
229 6.7 Hz, 1H), 7.20 (m, 2H), 7.14 (d, *J* = 7.8 Hz, 1H), 2.36 (s, 3H) ppm.

230
231
232 4.3.5 2-(4-((6-bromo-3-nitro-2-oxo-2H-chromen-4-yl)amino)phenyl)-2-methylpropanenitrile (**6e**)

233 Yield: 95%. ESI-MS: Calcd for C₁₉H₁₄BrN₃O₄ [M-H]⁻: 426.02, 429.01, Found 426.1, 428.1. ¹H
234 NMR (400 MHz, DMSO) δ 10.37 (s, 1H), 8.33 (d, *J* = 8.9 Hz, 1H), 7.85 (d, *J* = 1.8 Hz, 1H), 7.75
235 (dd, *J* = 8.7, 1.7 Hz, 1H), 7.51 (d, *J* = 8.6 Hz, 2H), 7.25 (d, *J* = 8.5 Hz, 2H), 1.69 (s, 6H) ppm.

236
237 4.4 General procedure for the synthesis of **7a** - **7e**

238 A 250 mL 3-necked round-bottom flask was charged with a solution of iron (166.2 mmol) and
239 NH₄Cl (50 mmol) in 3:1 ethanol: water (70 mL), heated to 100 °C, charged with **6** (6 g, 16.62
240 mmol) in several batches and stirred for 5 h. The reaction mixture was cooled, and added into a
241 solution of Na₂CO₃. The separated solid was filtered off and extracted with ethyl acetate. The
242 combined organic layers were dried over sodium sulfate, filtered and concentrated in vacuo.

243
244 4.4.1 3-amino-6-bromo-4-(phenylamino)-2H-chromen-2-one (**7a**)

245 Yield: 59%. ESI-MS: Calcd for C₁₅H₁₁BrN₂O₂ [M+H]⁺: 331.00, 333.00, Found 330.6, 332.7. ¹H
246 NMR (400 MHz, CDCl₃): δ 7.57 (d, *J* = 1.6 Hz, 1H), 7.41 (dd, *J* = 2.0, 8.8 Hz, 1H), 7.29 (m, 2H),
247 7.22 (d, *J* = 8.4 Hz, 1H), 6.94 (t, *J* = 14.4, 7.2 Hz, 1H), 7.21 (d, *J* = 8.0 Hz, 2H), 5.50 (s, 1H), 4.13
248 (br, 2H) ppm.

249
250 4.4.2 2-(4-((3-amino-6-bromo-2-oxo-2H-chromen-4-yl)amino)phenyl)acetonitrile (**7b**)

251 Yield: 63 %. ESI-MS: Calcd for C₁₇H₁₂BrN₃O₂ [M+Na]⁺: 392.01, 394.01, Found 389.9, 391.9.
252 ¹H NMR (400 MHz, CDCl₃): δ 7.53 (d, *J* = 2.4 Hz, 1H), 7.43 (dd, *J* = 8.8, 2.4 Hz, 1H), 7.22 (m,
253 4H), 6.71 (d, *J* = 8.4 Hz, 1H), 5.44 (s, 1H), 4.13 (br, 2H), 3.75 (s, 2H) ppm.

254
255 4.4.3 3-amino-6-bromo-4-((4-methoxyphenyl)amino)-2H-chromen-2-one (**7c**)

256 Yield: 54%. ESI-MS: Calcd for C₁₆H₁₃BrN₂O₃ [M+H]⁺: 361.01, 363.01, Found 360.6, 363.7. ¹H
257 NMR (400 MHz, CDCl₃): δ 7.55 (s, 1H), 7.42 (d, *J* = 8.8 Hz, 1H), 7.23 (d, *J* = 8.6 Hz, 1H), 6.86
258 (d, *J* = 8.1 Hz, 2H), 6.73 (d, *J* = 8.2 Hz, 2H), 5.41 (s, 1H), 3.79 (s, 3H), 1.26 (s, 2H) ppm.

4.4.4 3-amino-6-bromo-4-(o-tolylamino)-2H-chromen-2-one (**7d**)

Yield: 52%. ESI-MS: Calcd for C₁₆H₁₃BrN₂O₂ [M+ K]⁺: 383.02, 385.01, Found 383.2, 384.5. ¹H NMR (400 MHz, DMSO) δ 7.95 (s, 1H), 7.42 (dd, *J* = 8.7, 1.6 Hz, 1H), 7.32 (d, *J* = 7.7 Hz, 2H), 7.17 (d, *J* = 7.3 Hz, 1H), 6.95 (t, *J* = 7.5 Hz, 1H), 6.89 (s, 1H), 6.78 (t, *J* = 7.3 Hz, 1H), 6.31 (d, *J* = 7.9 Hz, 1H), 5.27 (s, 1H), 2.37 (s, 3H) ppm.

4.4.5 2-(4-((3-amino-6-bromo-2-oxo-2H-chromen-4-yl)amino)phenyl)-2-methylpropanenitrile (**7e**)

Yield: 61%. ESI-MS: Calcd for C₁₉H₁₆BrN₃O₂ [M+ Na]⁺: 420.04, 422.04, Found 420.2, 422.5. ¹H NMR (400 MHz, d⁶-DMSO): δ 7.96 (s, 1H), 7.55 (d, *J* = 2.0 Hz, 1H), 7.45 (dd, *J* = 1.8 Hz, 8.6 Hz, 1H), 7.35 (s, 1H), 7.32 (d, *J* = 9.2 Hz, 2H), 6.67 (d, *J* = 8.8 Hz, 2H), 5.24 (s, 2H), 1.63 (s, 6H) ppm.

4.5 General procedure for the synthesis of **8a** - **8e**

Compound **7** (9 mmol) and 1, 1'-carbonyldiimidazole (45 mmol) in acetic acid (50 mL) were stirred for 12 h at 120 °C. The reaction mixture was cooled and concentrated in vacuo to remove most of the acetic acid. The residual solution was added into water. The precipitate was filtered and extracted with ethyl acetate and then it was purified by silica gel chromatography.

4.5.1 8-bromo-1-phenyl-1,3-dihydrochromeno[3,4-*d*]imidazole-2,4-dione (**8a**)

Yield: 74%. ESI-MS: Calcd for C₁₆H₉BrN₂O₃ [M+ H]⁺: 356.98, 358.98, Found 355.1, 357.1. ¹H NMR (400 MHz, d⁶-DMSO): δ 12.12 (s, 1H), 7.68 (d, *J* = 3.2 Hz, 3H), 7.61 (m, 3H), 7.46 (d, *J* = 8.8 Hz, 1H), 6.62 (d, *J* = 2.0 Hz, 1H) ppm.

4.5.2 2-(4-(8-bromo-2,4-dioxo-2,3-dihydrochromeno[3,4-*d*]imidazol-1(4H)-yl)phenyl)acetonitrile (**8b**)

Yield: 70%. ESI-MS: Calcd for C₁₈H₁₀BrN₃O₃ [M+ H]⁺: 395.99, 397.99 Found 393.9, 395.9. ¹H NMR (400 MHz, d⁶-DMSO): δ 7.76 (s, 4H), 7.65 (m, 1H), 7.50 (d, *J* = 8.8 Hz, 1H), 6.62 (d, *J* = 2.0 Hz, 1H), 4.30 (s, 2H) ppm

4.5.3 8-bromo-1-(4-methoxyphenyl)-1,3-dihydrochromeno[3,4-*d*]imidazole-2,4-dione (**8c**)

Yield: 45%. ESI-MS: Calcd for C₁₇H₁₁BrN₂O₄ [M+ H]⁺: 386.99, 388.99 Found 385.0, 387.0. ¹H NMR (400 MHz, d⁶-DMSO): δ 7.62 (m, 3H), 7.47 (d, *J* = 8.9 Hz, 1H), 7.29 (d, *J* = 8.7 Hz, 2H), 6.67 (d, *J* = 2.0 Hz, 1H), 3.92 (s, 3H) ppm. ESI-MS: 385, 387 [M+H]⁺.

4.5.4 8-bromo-1-(o-tolyl)-1,3-dihydrochromeno[3,4-*d*]imidazole-2,4-dione (**8d**)

Yield: 62%. ESI-MS: Calcd for C₁₇H₁₁BrN₂O₃ [M+ H]⁺: 370.00, 371.99 Found 369.1, 371.1. ¹H NMR (400 MHz, d⁶-DMSO): δ 7.65 (m, 5H), 7.49 (d, *J* = 8.8 Hz, 1H), 6.45 (d, *J* = 1.4 Hz, 1H), 1.98 (s, 3H) ppm.

4.5.5

2-(4-(8-bromo-2,4-dioxo-2,3-dihydrochromeno[3,4-*d*]imidazol-1(4H)-yl)phenyl)-2-methylpropane nitrile (**8e**)

Yield: 56%. ESI-MS: Calcd for C₂₀H₁₄BrN₃O₃ [M+ Na]⁺: 446.02, 448.02 Found 444.3, 446.3.

¹H NMR (400 MHz, d⁶-DMSO): δ 7.93 (d, *J* = 8.8 Hz, 2H), 7.81 (d, *J* = 8.4 Hz, 2H), 7.63 (dd, *J* = 2.0 Hz, 8.8 Hz, 1H), 7.49 (d, *J* = 8.8 Hz, 1H), 6.41 (d, *J* = 2.4 Hz, 1H), 1.83 (s, 6H) ppm.

4.6 General procedure for the synthesis of **9-35**

A mixture of **8** (0.1 mmol), boric acid (0.11 mmol), PdCl₂ (dppf) CH₂Cl₂ (0.05 mmol) and K₂CO₃ (0.3 mmol) in dioxane/water (3/1 mL) was bubbled with nitrogen for 10 minutes, and heated in a sealed tube at 50 °C for 5 h. After cooling the reaction mixture was partitioned with water. The aqueous layer was further extracted with ethyl acetate. The organic extract was washed with brine, dried over sodium sulfate, and it was purified by silica gel chromatography.

4.6.1 1,8-diphenyl-1,3-dihydrochromeno[3,4-*d*]imidazole-2,4-dione (**9**)

Yield: 44%. ESI-MS: Calcd for C₂₂H₁₄N₂O₃ [M+ H]⁺: 355.10 Found 353.2. ¹H NMR (400 MHz, CDCl₃) δ 7.71 (s, 3H), 7.61 (d, *J* = 8.6 Hz, 1H), 7.49 (d, *J* = 8.4 Hz, 3H), 7.32 (m, 3H), 7.19 (d, *J* = 7.2 Hz, 2H), 6.84 (s, 1H) ppm.

4.6.2 1-phenyl-8-(pyridin-3-yl)-1,3-dihydrochromeno[3,4-*d*]imidazole-2,4-dione (**10**)

Yield: 43%. ESI-MS: Calcd for C₂₁H₁₃N₃O₃ [M+ Na]⁺: 378.10 Found 376.0. ¹H NMR (400 MHz, CDCl₃) δ 8.54 (d, *J* = 3.7 Hz, 1H), 8.41 (d, *J* = 1.8 Hz, 1H), 7.73 (dt, *J* = 4.2 Hz, 5.4 Hz, 3H, 3H), 7.60 (dd, *J* = 8.6, 2.1 Hz, 1H), 7.55 (m, 2H), 7.49 (dd, *J* = 7.7, 1.7 Hz, 2H), 7.29 (dd, *J* = 8.0, 4.9 Hz, 1H), 6.82 (d, *J* = 2.0 Hz, 1H) ppm.

4.6.3 8-(3-aminophenyl)-1-phenyl-1,3-dihydrochromeno[3,4-*d*]imidazole-2,4-dione (**11**)

Yield: 57%. ESI-MS: Calcd for C₂₂H₁₅N₃O₃ [M+ H]⁺: 370.11 Found 368.3. ¹H NMR (400 MHz, DMSO) δ 7.76 (d, *J* = 5.7 Hz, 5H), 7.62 (d, *J* = 8.6 Hz, 1H), 7.54 (d, *J* = 8.6 Hz, 1H), 6.97 (t, *J* = 7.7 Hz, 1H), 6.72 (s, 1H), 6.50 (d, *J* = 8.0 Hz, 1H), 6.47 (s, 1H), 6.28 (d, *J* = 7.5 Hz, 1H), 5.11 (s, 2H) ppm.

4.6.4 1-phenyl-8-(*o*-tolyl)-1,3-dihydrochromeno[3,4-*d*]imidazole-2,4-dione (**12**)

Yield: 55%. ESI-MS: Calcd for C₂₃H₁₆N₂O₃ [M+ Na]⁺: 391.12 Found 389.0. ¹H NMR (400 MHz, CDCl₃) δ 7.62 (s, 3H), 7.49 (d, *J* = 8.6 Hz, 1H), 7.43 (s, 2H), 7.35 (d, *J* = 8.6 Hz, 1H), 7.18 (m, 3H), 7.00 (d, *J* = 7.2 Hz, 1H), 6.66 (s, 1H), 2.38 (s, 3H) ppm.

4.6.5 8-(4-hydroxyphenyl)-1-phenyl-1,3-dihydrochromeno[3,4-*d*]imidazole-2,4-dione (**13**)

Yield: 52%. ESI-MS: Calcd for C₂₂H₁₄N₂O₄ [M- H]⁻: 369.10 Found 367.1. ¹H NMR (400 MHz, DMSO) δ 9.59 (s, 1H), 7.75 (d, *J* = 3.1 Hz, 5H), 7.65 (dd, *J* = 8.7, 2.2 Hz, 1H), 7.50 (d, *J* = 8.7 Hz, 1H), 7.01 (d, *J* = 8.6 Hz, 2H), 6.71 (d, *J* = 8.6 Hz, 2H), 6.65 (d, *J* = 2.0 Hz, 1H) ppm.

4.6.6

N-(3-(2,4-dioxo-1-phenyl-1,2,3,4-tetrahydrochromeno[3,4-*d*]imidazol-8-yl)phenyl)-4-fluorobenzenesulfonamide (**14**)

Yield: 56%. ESI-MS: Calcd for C₂₈H₁₈FN₃O₅S [M- H]⁻: 526.10 Found 524.3. ¹H NMR (400 MHz, DMSO) δ 10.40 (s, 1H), 7.81 (m, 2H), 7.74 (s, 5H), 7.58 (s, 2H), 7.40 (t, *J* = 8.3 Hz, 2H), 7.22 (t, *J* = 7.8 Hz, 1H), 7.10 (s, 1H), 7.00 (d, *J* = 8.0 Hz, 1H), 6.80 (d, *J* = 7.7 Hz, 1H), 6.71 (s,

1H) ppm.

4.6.7 1-phenyl-8-(pyridin-4-yl)-1,3-dihydrochromeno[3,4-d]imidazole-2,4-dione (15)

Yield: 57%. ESI-MS: Calcd for $C_{21}H_{13}N_3O_3$ $[M+H]^+$: 356.10 Found 354.0. 1H NMR (400 MHz, $CDCl_3$) δ 8.57 (d, J = 4.8 Hz, 2H), 7.74 (m, 3H), 7.66 (dd, J = 8.7, 1.9 Hz, 1H), 7.52 (m, 3H), 7.09 (d, J = 5.4 Hz, 2H), 6.87 (s, 1H) ppm.

4.6.8 1-phenyl-8-(quinolin-3-yl)-1,3-dihydrochromeno[3,4-d]imidazole-2,4-dione (16)

Yield: 40%. ESI-MS: Calcd for $C_{25}H_{15}N_3O_3$ $[M+Na]^+$: 428.11 Found 426.1. 1H NMR (400 MHz, $CDCl_3$) δ 8.72 (d, J = 2.2 Hz, 1H), 8.11 (d, J = 8.4 Hz, 1H), 8.00 (d, J = 1.9 Hz, 1H), 7.75 (m, 6H), 7.59 (t, J = 8.0 Hz, 2H), 7.52 (m, 2H), 6.94 (d, J = 2.0 Hz, 1H) ppm.

4.6.9

2-(4-(2,4-dioxo-8-(o-tolyl)-2,3-dihydrochromeno[3,4-d]imidazol-1(4H)-yl)phenyl)acetonitrile (17)

Yield: 52%. ESI-MS: Calcd for $C_{25}H_{17}N_3O_3$ $[M-H]^-$: 406.13 Found 404.2. 1H NMR (400 MHz, $CDCl_3$) δ 7.63 (d, J = 8.1 Hz, 2H), 7.49 (dd, J = 8.2, 6.0 Hz, 3H), 7.37 (dd, J = 8.6, 2.0 Hz, 1H), 7.19 (m, 3H), 7.00 (d, J = 7.1 Hz, 1H), 6.65 (d, J = 1.9 Hz, 1H), 3.88 (s, 2H), 2.37 (s, 3H) ppm.

4.6.10

N-(3-(1-(4-(cyanomethyl)phenyl)-2,4-dioxo-1,2,3,4-tetrahydrochromeno[3,4-d]imidazol-8-yl)phenyl)-4-fluorobenzenesulfonamide (18)

Yield: 43%. ESI-MS: Calcd for $C_{30}H_{19}FN_4O_5S$ $[M-H]^-$: 565.11 Found 563.2. 1H NMR (400 MHz, DMSO) δ 10.43 (s, 1H), 7.78 (m, 6H), 7.61 (s, 2H), 7.39 (t, J = 8.8 Hz, 2H), 7.25 (t, J = 7.9 Hz, 1H), 7.16 (s, 1H), 6.94 (d, J = 8.1 Hz, 1H), 6.79 (d, J = 7.9 Hz, 1H), 6.74 (s, 1H), 4.25 (s, 2H) ppm.

4.6.11

2-(4-(8-(3-aminophenyl)-2,4-dioxo-2,3-dihydrochromeno[3,4-d]imidazol-1(4H)-yl)phenyl)acetonitrile (19)

Yield: 55%. ESI-MS: Calcd for $C_{24}H_{16}N_4O_3$ $[M+H]^+$: 409.12 Found 407.3. 1H NMR (400 MHz, $CDCl_3$) δ 7.72 (d, J = 7.9 Hz, 2H), 7.56 (dd, J = 15.8, 8.9 Hz, 3H), 7.46 (d, J = 8.6 Hz, 1H), 7.15 (t, J = 7.8 Hz, 1H), 6.76 (s, 1H), 6.63 (d, J = 8.5 Hz, 1H), 6.57 (d, J = 7.5 Hz, 1H), 6.50 (s, 1H), 3.96 (s, 2H), 3.79 (s, 2H) ppm.

4.6.12

2-(4-(2,4-dioxo-8-phenyl)-2,3-dihydrochromeno[3,4-d]imidazol-1(4H)-yl)phenyl)acetonitrile (20)

Yield: 55%. ESI-MS: Calcd for $C_{24}H_{15}N_3O_3$ $[M-H]^-$: 392.11 Found 390.2. 1H NMR (400 MHz, $CDCl_3$) δ 7.72 (d, J = 8.2 Hz, 2H), 7.62 (dd, J = 8.7, 2.1 Hz, 1H), 7.56 (d, J = 8.2 Hz, 2H), 7.50 (d, J = 8.6 Hz, 1H), 7.39 (t, J = 7.4 Hz, 2H), 7.32 (t, J = 7.3 Hz, 1H), 7.20 (d, J = 7.2 Hz, 2H), 6.79 (d, J = 2.0 Hz, 1H), 3.95 (s, 2H) ppm.

4.6.13

2-(4-(8-(4-hydroxyphenyl)-2,4-dioxo-2,3-dihydrochromeno[3,4-d]imidazol-1(4H)-yl)phenyl)acetonitrile (21)

Yield: 55%. ESI-MS: Calcd for $C_{24}H_{15}N_3O_4$ $[M-H]^-$: 308.11 Found 306.2. 1H NMR (400 MHz, d^6 -DMSO): δ 9.64 (s, 1H), 7.79 (m, 4H), 7.70 (dd, $J = 2.0$ Hz, 8.8 Hz, 1H), 7.52 (d, $J = 8.8$ Hz, 1H), 7.05 (d, $J = 8.8$ Hz, 2H), 6.79 (d, $J = 8.4$ Hz, 2H), 6.67 (d, $J = 2.0$ Hz, 1H), 4.30 (s, 2H) ppm.

4.6.14

2-(4-(2,4-dioxo-8-(pyridin-3-yl)-2,3-dihydrochromeno[3,4-d]imidazol-1(4H)-yl)phenyl)acetonitrile (22)

Yield: 51%. ESI-MS: Calcd for $C_{23}H_{14}N_4O_3$ $[M+H]^+$: 395.11 Found 393.1. 1H NMR (400 MHz, $CDCl_3$): δ 8.55 (s, 1H), 8.45 (s, 1H), 7.23 (d, $J = 8.4$ Hz, 2H), 7.59 (d, $J = 8.4$ Hz, 3H), 7.51 (t, $J = 9.6$ Hz, 2H), 7.35 (m, 1H), 6.78 (d, $J = 1.6$ Hz, 1H), 3.99 (s, 2H) ppm.

4.6.15

2-(4-(8-(4-(methylsulfonyl)phenyl)-2,4-dioxo-2,3-dihydrochromeno[3,4-d]imidazol-1(4H)-yl)phenyl)acetonitrile (23)

Yield: 49%. ESI-MS: Calcd for $C_{25}H_{17}N_3O_5S$ $[M+H]^+$: 472.09 Found 470.2. 1H NMR (400 MHz, $CDCl_3$) δ 7.96 (d, $J = 8.2$ Hz, 2H), 7.73 (d, $J = 7.7$ Hz, 2H), 7.64 (d, $J = 7.3$ Hz, 1H), 7.56 (m, 3H), 7.37 (d, $J = 8.2$ Hz, 2H), 6.80 (s, 1H), 3.96 (s, 2H), 3.08 (s, 3H) ppm.

4.6.16

2-(4-(2,4-dioxo-8-(pyridin-4-yl)-2,3-dihydrochromeno[3,4-d]imidazol-1(4H)-yl)phenyl)acetonitrile (24)

Yield: 58%. ESI-MS: Calcd for $C_{23}H_{14}N_4O_3$ $[M+H]^+$: 395.11 Found 393.0. 1H NMR (400 MHz, $CDCl_3$) δ 8.62 (s, 2H), 7.74 (d, $J = 8.2$ Hz, 2H), 7.67 (dd, $J = 8.6, 2.1$ Hz, 1H), 7.56 (d, $J = 7.6$ Hz, 3H), 7.11 (d, $J = 2.8$ Hz, 2H), 6.84 (d, $J = 2.0$ Hz, 1H), 3.97 (s, 2H) ppm.

4.6.17

2-(4-(2,4-dioxo-8-(quinolin-3-yl)-2,3-dihydrochromeno[3,4-d]imidazol-1(4H)-yl)phenyl)acetonitrile (25)

Yield: 52%. ESI-MS: Calcd for $C_{27}H_{16}N_4O_3$ $[M+H]^+$: 445.12 Found 443.3. 1H NMR (400 MHz, $CDCl_3$): δ 8.72 (s, 1H), 8.10 (d, $J = 8.4$ Hz, 1H), 8.01 (s, 1H), 7.84 (d, $J = 8.0$ Hz, 1H), 7.75 (m, 4H), 7.60 (m, 4H), 7.27 (s, 1H), 6.92 (s, 1H), 3.99 (s, 2H) ppm.

4.6.18 1-(4-methoxyphenyl)-8-(pyridin-3-yl)-1,3-dihydrochromeno[3,4-d]imidazole-2,4-dione (26)

Yield: 47%. ESI-MS: Calcd for $C_{22}H_{15}N_3O_4$ $[M+H]^+$: 386.11 Found 384.0. 1H NMR (400 MHz, $CDCl_3$) δ 8.56 (d, $J = 4.2$ Hz, 1H), 8.50 (s, 1H), 7.56 (m, 3H), 7.39 (d, $J = 8.6$ Hz, 2H), 7.30 (m, 1H), 7.18 (d, $J = 8.6$ Hz, 2H), 6.92 (s, 1H), 3.96 (d, $J = 7.8$ Hz, 3H) ppm.

4.6.19 1-(4-methoxyphenyl)-8-(pyridin-4-yl)-1,3-dihydrochromeno[3,4-d]imidazole-2,4-dione (27)

Yield: 43%. ESI-MS: Calcd for $C_{22}H_{15}N_3O_4$ $[M+H]^+$: 386.11 Found 384.0. 1H NMR (400 MHz, $CDCl_3$) δ 8.59 (s, 2H), 7.66 (d, $J = 8.2$ Hz, 1H), 7.54 (d, $J = 8.6$ Hz, 1H), 7.40 (d, $J = 8.4$ Hz, 2H), 7.19 (m, 4H), 6.94 (s, 1H), 3.97 (s, 3H) ppm.

4.6.20 1-(4-methoxyphenyl)-8-(quinolin-3-yl)-1,3-dihydrochromeno[3,4-d]imidazole-2,4-dione (28)

Yield: 53%. ESI-MS: Calcd for $C_{26}H_{17}N_3O_4$ $[M+H]^+$: 436.12 Found 334.1. 1H NMR (400 MHz, $CDCl_3$) δ 8.85 (s, 2H), 7.79 (m, 4H), 7.60 (d, J = 8.1 Hz, 1H), 7.42 (d, J = 7.9 Hz, 3H), 7.22 (d, J = 7.0 Hz, 2H), 7.03 (s, 1H), 3.97 (s, 3H) ppm.

4.6.21 8-(quinolin-3-yl)-1-(o-tolyl)-1,3-dihydrochromeno[3,4-d]imidazole-2,4-dione (**29**)

Yield: 55%. ESI-MS: Calcd for $C_{26}H_{17}N_3O_3$ $[M+H]^+$: 420.13 Found 418.2. 1H NMR (400 MHz, $CDCl_3$) δ 8.65 (s, 1H), 8.05 (d, J = 8.2 Hz, 1H), 7.92 (s, 1H), 7.73 (d, J = 8.0 Hz, 1H), 7.66 (m, 3H), 7.59 (d, J = 7.3 Hz, 1H), 7.49 (m, 3H), 7.36 (d, J = 7.6 Hz, 1H), 6.76 (s, 1H), 2.01 (s, 3H) ppm.

4.6.22 8-(pyridin-3-yl)-1-(o-tolyl)-1,3-dihydrochromeno[3,4-d]imidazole-2,4-dione (**30**)

Yield: 51%. ESI-MS: Calcd for $C_{22}H_{15}N_3O_3$ $[M+H]^+$: 370.11 Found 368.3. 1H NMR (400 MHz, $CDCl_3$) δ 8.48 (s, 1H), 8.34 (s, 1H), 7.51 (m, 6H), 7.33 (d, J = 7.7 Hz, 1H), 7.22 (s, 1H), 6.65 (d, J = 1.4 Hz, 1H), 1.98 (d, J = 5.8 Hz, 3H) ppm.

4.6.23 8-(pyridin-4-yl)-1-(o-tolyl)-1,3-dihydrochromeno[3,4-d]imidazole-2,4-dione (**31**)

Yield: 46%. ESI-MS: Calcd for $C_{22}H_{15}N_3O_3$ $[M+H]^+$: 370.11 Found 368.3. 1H NMR (400 MHz, $CDCl_3$) δ 8.57 (s, 2H), 7.66 (dd, J = 11.3, 4.3 Hz, 2H), 7.56 (dd, J = 12.8, 7.7 Hz, 3H), 7.42 (d, J = 7.6 Hz, 1H), 7.08 (d, J = 4.2 Hz, 2H), 6.77 (d, J = 1.8 Hz, 1H), 2.06 (s, 3H) ppm.

4.6.24

2-(4-(2,4-dioxo-8-(pyridin-3-yl)-2,3-dihydrochromeno[3,4-d]imidazol-1(4H)-yl)phenyl)-2-methylpropanenitrile (**32**)

Yield: 60%. ESI-MS: Calcd for $C_{25}H_{18}N_4O_3$ $[M+Na]^+$: 445.14 Found 443.1. 1H NMR (400 MHz, $CDCl_3$) δ 8.54 (s, 1H), 8.43 (s, 1H), 7.85 (d, J = 8.1 Hz, 2H), 7.55 (m, 4H), 7.28 (s, 2H), 6.69 (s, 1H), 1.87 (s, 6H) ppm.

4.6.25

2-(4-(2,4-dioxo-8-(pyridin-4-yl)-2,3-dihydrochromeno[3,4-d]imidazol-1(4H)-yl)phenyl)-2-methylpropanenitrile (**33**)

Yield: 56%. ESI-MS: Calcd for $C_{25}H_{18}N_4O_3$ $[M+Na]^+$: 445.14 Found 443.1. 1H NMR (400 MHz, $CDCl_3$) δ 8.56 (d, J = 4.8 Hz, 2H), 7.87 (d, J = 8.2 Hz, 2H), 7.67 (m, 2H), 7.56 (t, J = 8.5 Hz, 3H), 7.08 (d, J = 4.5 Hz, 2H), 6.78 (s, 1H), 1.88 (s, 6H) ppm.

4.6.26

2-(4-(2,4-dioxo-8-(quinolin-3-yl)-2,3-dihydrochromeno[3,4-d]imidazol-1(4H)-yl)phenyl)-2-methylpropanenitrile (**34**)

Yield: 48%. ESI-MS: Calcd for $C_{29}H_{20}N_4O_3$ $[M+Na]^+$: 495.15 Found 493.2. 1H NMR (400 MHz, d^6 -DMSO): δ 8.71 (d, J = 2.0 Hz, 1H), 8.28 (s, 1H), 7.98 (m, 5H), 7.86 (d, J = 8.4 Hz, 2H), 7.78 (t, J = 7.8 Hz, 1H), 7.71 (d, J = 8.4 Hz, 1H), 7.65 (t, J = 7.4 Hz, 1H), 6.82 (d, J = 1.6 Hz, 1H), 2.65 (s, 6H) ppm.

4.7 Procedure for the synthesis of **35a**

A mixture of compound **8e** (1 mmol), CH_3I (1 mmol), and potassium tert-butanolate (3 mmol) in DMF was stirred at 0 °C for 20 minutes. The reaction mixture was partitioned with water. Then

the aqueous layer was further extracted with ethyl acetate. The organic extract was washed with brine, dried over sodium sulfate, and it was purified by silica gel chromatography.

2-(4-(8-bromo-2-(tert-butoxy)-2,3-dimethyl-4-oxo-2,3-dihydrochromeno[3,4-d]imidazol-1(4H)-yl)phenyl)-2-methylpropanenitrile (35a)

Yield: 7%. ESI-MS: Calcd for $C_{26}H_{28}BrN_3O_3$ $[M+H]^+$: 510.13, 512.13 Found 510.2, 512.2 1H NMR (400 MHz, DMSO): δ 7.55 (d, J = 8.3 Hz, 2H), 7.44 (dd, J = 8.8, 2.3 Hz, 1H), 7.37 (d, J = 2.3 Hz, 1H), 7.29 (d, J = 8.2 Hz, 2H), 6.84 (d, J = 8.9 Hz, 1H), 3.49 (s, 3H), 2.18 (s, 3H), 1.66 (s, 6H), 1.23 (s, 9H) ppm. ESI-MS: 510.2, 512.2 $[M+H]^+$.

4.7.1

2-(4-(2-(tert-butoxy)-2,3-dimethyl-4-oxo-8-(quinolin-3-yl)-2,3-dihydrochromeno[3,4-d]imidazol-1(4H)-yl)phenyl)-2-methylpropanenitrile (35)

Yield: 55%. ESI-MS: Calcd for $C_{35}H_{34}N_4O_3$ $[M+H]^+$: 559.26 Found 559.3. 1H NMR (400 MHz, $CDCl_3$): δ 9.03 (s, 1H), 8.16 (s, 1H), 8.11 (d, J = 8.4 Hz, 1H), 7.84 (d, J = 8.1 Hz, 1H), 7.71 (t, J = 7.6 Hz, 1H), 7.64 (d, J = 8.5 Hz, 1H), 7.57 (t, J = 7.5 Hz, 1H), 7.52 (s, 1H), 7.46 (d, J = 8.2 Hz, 2H), 7.16 (d, J = 8.1 Hz, 2H), 6.90 (d, J = 8.6 Hz, 1H), 3.64 (s, 3H), 2.36 (s, 3H), 1.68 (s, 6H), 1.36 (s, 9H) ppm. ^{13}C NMR (400 MHz, DMSO): δ 161.71, 157.12, 149.03, 146.46, 144.82, 131.86, 131.62, 131.21, 130.16, 129.23, 128.86, 128.61, 128.33, 128.17, 127.90, 127.67, 127.01, 125.93, 124.27, 119.72, 111.69, 78.85, 55.40, 36.45, 28.05, 27.60, 13.64 ppm. Mp = 205- 207 °C.

4.8. Biological evaluation

4.8.1 Cell Culture and reagents preparation

Mouse colorectal carcinoma CT26 cell, human colorectal carcinoma HCT116 and SW620 cells, human breast carcinoma MCF-7 cell, lung carcinoma A549 cell and human hepatocytes LO2 cell were obtained from the American Type Culture Collection (ATCC, Rockville, MD, USA). Cells were propagated in DMEM or RPMI 1350 media containing 10% fetal bovine serum (FBS; Gibco, Auckland, N. Z.) and 1% antibiotics (penicillin and streptomycin) in 5% CO_2 at 37 °C.

4.8.2 MTT Assay

The cell viability was performed by MTT assay. Briefly, cells were seeded in 96-well plates (2000 - 3000 cells/plate). After 24h incubation, the cells were treated with various concentrations of compounds. After treatment for 72 h, a volume of 20 μ L of MTT solution (5 mg/mL) was added to each well and incubated for additional 2 - 4 h incubation at 37 °C. Then the medium was discarded, and the formazan salt was dissolved with 150 μ L DMSO for 15 - 20 minutes. The absorbance of each well was measured at 570 nm using a Spectra MAX M5 microplate spectrophotometer (Molecular Devices, CA, USA), and the median inhibitory concentration (IC_{50}) values were calculated. Three replicate wells were used for each analysis. The results were obtained from three separate experiments.

4.8.3 Colony Formation Assay

To test the survival of cells treated with compound **35**, HCT116 cells (500 cells/well) were plated in a 6-well plate and incubated overnight at 37 °C, followed by various concentrations of **35** treatment (0 - 5.0 μ M) for 15 days with fresh medium. Finally, the cells were washed with cold

phosphate-buffered saline (PBS) for twice. Colonies were fixed with 4% paraformaldehyde and stained with a 0.5% crystal violet solution for 15 minutes, and the colonies (>50 cells) were counted under microscope. The data are expressed from three independent experiments. The number of colonies in treated cultures was expressed as a percentage of the control cultures.

4.8.4 Flow Cytometry

The cell cycle distribution of SW620 cells after 24 h exposure to various concentrations of **35** was monitored by flow cytometric analysis using a FACScan flow cytometer (Becton Dickinson USA). Briefly, SW620 cells (2×10^5 cells/well) were seeded in 6-well plates and incubated overnight at 37 °C. After 24 h treatment, the cells were harvested and washed with cold PBS twice, and fixed with 75% alcohol at 4 °C. 24h later, cells were stained with propidium iodide (PI) and analyzed by flow cytometry. Then data were analyzed with FlowJo software.

4.8.5 Apoptotic Assay

To further confirm the apoptosis inducing effect of compound **35**, Annexin V-FITC apoptosis detection kit was used. HCT116 cells (2×10^5 cells/well) were seeded in 6-well plates and incubated overnight at 37 °C. After 24 h exposure to various concentrations of **35**, cells were harvested and washed with cold PBS twice. After centrifugation, cells were stained with Annexin V-FITC and PI, and then analyzed with FCM (Becton- Dickinson, USA). Then data were analyzed with FlowJo software.

4.8.6 Wound-healing migration assay

The inhibition of tumor cell migration by compound **35** was determined by wound-healing migration assay. MCF-7 cells were seeded onto 6-well plates, when grew to 80 - 90% confluence and then the “wound” were scraped by a 0.1 mL sterile pipette tips. Fresh medium containing 10% fetal bovine serum and different concentrations of **35** were added. Cells were photographed after 0, 12, 24 h incubation at 37 °C, respectively.

4.8.7 Mice and tumor model

All animal experiments were approved and conducted by the Institutional Animal Care and Treatment Committee of Sichuan University in China. 100 mL tumor cell (HCT116) suspension containing 1×10^7 cells were injected subcutaneously into the right flank of the seven-week-old female BALB/c athymic nude mice. After one week, when average tumor volume reached approximately 100 mm^3 , mice were separated into three groups randomly (8 mice per group), and received intraperitoneally injection (i.p.) of **35** at dose of 40 mg/kg, 80 mg/kg or vehicle, respectively once two days for 24 days. Tumor growth and body weight were measured every two days during the treatment. The tumor volumes were calculated according to the formula as follow: $\text{volume (mm}^3\text{)} = (L \times W^2) \times 0.5$, where L is the length and W is the width. Growth inhibition was calculated from the start of treatment by comparison of the mean change in tumor volume for vehicle and treated group as before.

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Fig. 1. A few of benzimidazole and imidazoquinoline derivatives with antitumor activities.

Fig. 2. The effect of compound 35 on cancer cells viability (A) Proliferation of HCT116, MCF-7, SW620, CT26, A549 and LO2 cells treated with various concentrations (0 - 20 μ M) of 35 and Cisplatin for 72 h, respectively. Cell viability was detected by MTT assay. The data are expressed as the means $\pm$ SD from three independent experiments. (B) and (C) The colony clusters were detected after a 20-day 35 treatment. Quantification is shown in the right panels. Images shown are representatives of three independent experiments. (** $p < 0.01$).

Fig. 3. Compound 35 induced G0/G1 cell-cycle arrest and apoptosis of SW620 cells. (A) SW620 cells were treated with various concentrations of 35 (0, 0.3125, 0.625, 1.25, 5.0 μ M) for 24 h. (B) SW620 cells were exposed to 35 at indicated doses for 24 h, and the level of apoptosis was evaluated using the Annexin V/PI dual-labeling technique, and analyzed by flow cytometry. Data shown are representative of three independent experiments.

Fig. 4. Compound 35 inhibited breast cancer cell MCF-7 migration *in vitro*.

Fig. 5. Effects of compound 35 on the growth of xenografts in nude mice *in vivo*. (A) HCT116 tumor-bearing nude mice were treated as described with vehicle, 35 at 40 and 80 mg/kg, the mean tumor volumes $\pm$ SD of five mice per every group. (B) After the tumor cell inoculation, the body weight of the 35 treatment and vehicle groups were counted, and there were no significant difference among the groups.

Tables

Table 1. *In Vitro* inhibition of tumor cell growth of compounds **9** - **34**.

Compd.	R ¹	R ²	IC ₅₀ (μM) ^a		
			U87-MG	PC-3	HCT116
9	C ₆ H ₅	C ₆ H ₅	>40	>40	—
10	C ₆ H ₅	pyridin-3-yl	>40	>40	—
11	C ₆ H ₅	C3-NH ₂ -C ₆ H ₄	>40	>40	—
12	C ₆ H ₅	2-Me-C ₆ H ₄	>40	—	—
13	C ₆ H ₅	4-OH-C ₆ H ₄	>40	—	—
14	C ₆ H ₅		>40	>40	>40
15	C ₆ H ₅	pyridin-4-yl	38.74 ± 1.64	-	-
16	C ₆ H ₅	quinolin-3-yl	35.6 ± 2.87	34.05 ± 3.43	31.5 ± 4.89
17	4-CH ₂ CN-C ₆ H ₄	2-Me-C ₆ H ₄	>40	>40	>40
18	4-CH ₂ CN-C ₆ H ₄		>40	>40	>40
19	4-CH ₂ CN-C ₆ H ₄	3-NH ₂ -C ₆ H ₄	>40	36.51 ± 3.32	-

20	4-CH ₂ CN-C ₆ H ₄	C ₆ H ₅	31.42 ± 5.46	35.31 ± 2.56	>40
21	4-CH ₂ CN-C ₆ H ₄	4-OH-C ₆ H ₄	29.59 ± 4.30	30.59 ± 3.21	30.13 ± 4.44
22	4-CH ₂ CN-C ₆ H ₄	pyridin-3-yl	>40	>40	29.37 ± 5.56
23	4-CH ₂ CN-C ₆ H ₄	4-(methylsulfonyl)phenyl	39.72 ± 0.22	29.61 ± 2.32	-
24	4-CH ₂ CN-C ₆ H ₄	pyridin-4-yl	36.42 ± 1.45	37.06 ± 3.50	37.2 ± 2.10
25	4-CH ₂ CN-C ₆ H ₄	quinolin-3-yl	34.9 ± 2.40	30.25 ± 3.60	28.40 ± 2.60
26	4-OMe-C ₆ H ₄	pyridin-3-yl	>40	>40	>40
27	4-OMe-C ₆ H ₄	pyridin-4-yl	>40	>40	>40
28	4-OMe-C ₆ H ₄	quinolin-3-yl	>40	>40	>40
29	2-Me-C ₆ H ₄	quinolin-3-yl	>40	>40	>40
30	2-Me-C ₆ H ₄	pyridin-3-yl	>40	>40	>40
31	2-Me-C ₆ H ₄	pyridin-4-yl	>40	>40	>40
32	4-C(CH ₃) ₂ -C ₆ H ₄	pyridin-3-yl	25.62 ± 3.80	38.20 ± 2.10	-
33	4-C(CH ₃) ₂ -C ₆ H ₄	pyridin-4-yl	28.55 ± 2.30	29.01 ± 4.60	-
34	4-C(CH ₃) ₂ -C ₆ H ₄	quinolin-3-yl	33.42 ± 2.50	19.76 ± 1.60	20.40 ± 3.60
Cisplatin	-	-	>20	>20	12.3

^a Inhibitory activity was assayed by exposure for 72 h to substances and expressed as concentration required to inhibit tumor cell proliferation by 50% (IC₅₀). SD, standard deviation (n = 3).

Table 2. *In vitro* Inhibition of tumor cell growth of compound **35**.

Compd.	R ²	IC ₅₀ (μM) ^a		
		U87-MG	OPC-9	HCT116
35	quinolin-3-yl	>31.0 ± 4.50	18.0 ± 2.40	1.40 ± 0.50
Cisplatin	-	>20	>20	12.3

^a Inhibitory activity was assayed by exposure for 72 h to substances and expressed as concentration required to inhibit tumor cell proliferation by 50% (IC₅₀). SD, standard deviation (n = 3).

Schemes

Scheme 1. Reagents and conditions: (i) acetic anhydride, pyridine, 100 °C, 3.5 h, 92%; (ii) AlCl₃, 150 °C, 3.5 h, 72%; (iii) diethyl carbonate, NaH, Toluene, 100 °C, 4 h, 71%; (iv) HNO₃, DCM, 0 °C, 2 h, 94%; (v) POCl₃, Et₃N, reflux, 1 h, 87%.

Scheme 2. Reagents and conditions: (i) DMF, r.t., 2h, 93-97%; (ii) Fe, ethanol, 75%, NH₄Cl, r.t., 4-5 h, 52-63 %; (iii) CDI, CH₃COOH, 2 h, 100 °C, 45-74%; (iv) K₂CO₃, PdCl₂ (dppf) CH₂Cl₂, 1,4-dioxane : H₂O=3:1, 65 °C, 4-5 h, 40-60%.

Scheme 3. Reagents and conditions: (i) CH₃I, Potassium tert-butanolate, DCM, 0 °C, 1 h, 7%; (ii) K₂CO₃, [PdCl₂ (dppf)] CH₂Cl₂, 1,4-dioxane : H₂O=3 : 1, 65 °C, 4-5 h, 55%.

Fig.1

Omeprazole

Telmisarta

Gsk121051A

NVP-BGT226

NVP-BEZ235

Fig.2

A

Comp.	IC ₅₀ (μM)					
	HCT116	MCF-7	SW620	CT26	A549	LO2
35	1.40 ± 0.5	1.70 ± 0.3	3.55 ± 1.3	3.50 ± 0.7	9.50 ± 1.8	37.0 ± 5.3
Cisplatin	12.30 ± 2.9	4.50 ± 1.8	12.40 ± 2.6	15.10 ± 3.6	9.97 ± 2.1	>20

B

C

Fig.3

Fig.4

Fig.5

Compound 35

HCT116 $IC_{50} = 1.40 \mu M$

MCF7 $IC_{50} = 1.70 \mu M$

Compound 35 (μM)

Compound 35 (μM)