Recovery of Cellular Traction in Three-Dimensional Nonlinear Hyperelastic Matrices

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Abstract

The traction exerted by a cell on the extra-cellular matrix (ECM) is critical to understanding and manipulating important biological processes such as stem cell differentiation, cancer cell metastasis, and embryonic morphogenesis. This traction is typically quantified through traction force microscopy (TFM). In TFM, the displacement of select markers inside the ECM is tracked, and is used in conjunction with an elasticity problem to reconstruct the traction field. Most applications of this technique thus far have assumed that the matrix behaves as a linear elastic solid that undergoes small deformation and infinitesimal strains. In this manuscript, we develop and implement a robust and efficient TFM methodology that overcomes these limitations by accounting for geometric and material nonlinearities in the ECM. We pose the TFM problem as an inverse problem and develop efficient adjoint-based minimization techniques to solve it. We test the effect of measurement noise on the proposed method, and examine the error incurred by not including nonlinear effects when solving the TFM problem. We present these results for *in-silico* traction fields that are applied to realistic geometric models of microglial and neuronal cells.

Keywords: Cell traction force microscopy, Material nonlinearity, Geometric nonlinearity, Stabilized finite element formulation, Inverse problem, Adjoint equations

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1. Introduction

The traction exerted by a cell on its surroundings (and by the surroundings on a cell) plays a critical role in cell development and migration, and is therefore important in understanding and manipulating important biological processes such as stem cell differentiation, cancer cell metastasis, and embryonic morphogenesis [1–3]. In order to quantitatively measure the traction field, several techniques have been developed. These include the use of wrinkling membranes [4], cantilever-based sensing devices [5, 6] and traction force microscopy (TFM)[7–9].

In TFM, the extracellular matrix (ECM) surrounding the cell is embedded with beads, and the motion of these beads in response to cellular tractions is tracked. Given this motion, and the mechanical behavior of the extracellular matrix, the traction at the cell-matrix interface is recovered [10]. When TFM is applied to a thick, two-dimensional, linear-elastic substrate, the problem of determining the tractions is further simplified by utilizing the Boussinesq solution [11] in the Fourier space. This technique is often referred to as Fourier Transform Traction Cytometry (FTTC)[8], and yields a rapid solution for the traction field. However, its application is limited to thick two-dimensional substrates undergoing small strains.

The extension of TFM to three dimensions is considered in [10]. This reference describes an approach to TFM for cells that are geometrically complex and are embedded in a three-dimensional matrix. This problem is solved by constructing a numerical Green’s function using the finite element method that connects the traction applied at a point on the cell-matrix interface to the displacement at a given bead location. Once this map is determined, a minimization problem is solved in order to determine the traction field that produces a bead displacement field that best matches the measured displacement field. A similar approach can also be found in [12].

A natural extension to the ideas described above would involve accounting for the nonlinear effects during the elastic deformation of the ECM. There are several sources of this nonlinear behavior:

1. When the strain in a solid exceeds a certain threshold (usually around 10%), the linear measure of strain (the infinitesimal strain tensor) is no longer appropriate, and other nonlinear measures that accurately measure the deformation of the material must be adopted [13–15]. We note that in TFM studies, where 30-70% strains are quite common [10, 16, 17], this level of strain is routinely exceeded.

2. When the displacement incurred by the solid is large, the change in the current configuration due to this displacement must be accounted for, and this introduces the so-called geometric nonlinearity to the problem.
3. Some materials display a nonlinear stress-strain response, and this leads to a nonlinear constitutive model. This is especially true for materials such as gelatin-agar co-gels [18], that are commonly used as tissue-mimicking phantom, or real biological gels [19].

For all these reasons, it is important to consider the nonlinear behavior of the ECM while computing the traction field exerted by the cell on the matrix.

We note that there have been some attempts at solving the 3D nonlinear TFM problem. In [20], the authors consider a nonlinear constitutive model, but do not account for nonlinearities in strain and in the geometrical description of the problem. Moreover, the applicability of their technique appears to be limited to a small range of traction magnitudes, and to a relatively small number of traction beads.

We also note that there are instances adopting a “forward computation method” (described as such by [21]) for determining the cell traction, in contrast to the inverse problem approach described above [9, 22–25]. In the forward method, the strain is obtained by differentiating the experimentally measured displacement field and the stress/traction fields are then determined by applying the constitutive equation. Clearly, the quality of the cellular tractions recovered using this approach depends heavily on the ability to measure the displacement field close to the cell-matrix interface [21]. This places a significant burden on the experimental set up. The inverse problem approach, on the other hand, can make use of all available bead displacement data, including those that are at some distance from the interface.

In this manuscript we present a new approach to solving the nonlinear TFM problem. We pose the problem as an inverse problem, where we are given the displacement of a finite number of beads, and a model for the mechanical response of the matrix, and we wish to determine the traction field that is consistent with these observations. We solve the inverse problem as a constrained minimization problem, where we seek to find the traction field that yields a displacement field that best matches measured bead displacements. This is done under the constraint that the predicted displacement field satisfies the equations of equilibrium for a nonlinear hyperelastic material. We solve this problem using gradient-based algorithms, and efficiently compute the gradient using an linearized adjoint equation.

75 We account for noise by regularizing the recovered traction field through a
 76 Tikhonov term. Similar methods for solving inverse problems have been used
 77 to determine the spatial distribution of material properties in different fields
 78 of mechanics and physics [26–44]. We test the performance of the proposed
 79 approach on synthetic data. We create geometric models of cells embedded
 80 in a matrix with known material behavior. Thereafter, we apply tractions at
 81 the cell-matrix interface and compute the displacement field in the matrix
 82 by solving a forward problem. We utilize a portion of the displacement field
 83 as the “measured” displacement field, and in some instances, add noise to
 84 it. Using this measured displacement in our algorithm, we determine the
 85 traction field at the interface. We also utilize this set-up to determine the
 86 effect of making the linear elastic assumption while solving the inverse prob-
 87 lem, when the actual response of the material is nonlinear. In the inverse
 88 problems literature, this error is referred to as model error. We conclude
 89 that for some problems the model error can be significant. Moreover, we
 90 note that the problem of recovering tractions is not limited to applications
 91 in biomechanics, and has also been studied in structural dynamics [45–47].
 92 In these studies also, the assumption of linear elasticity is invoked.

93 The organization of the remainder of this manuscript is as follows: In
 94 Section 2, we present the nonlinear forward elasticity problem that describes
 95 the response of a three-dimensional hydrogel to cellular traction. In Section
 96 3, we formulate the nonlinear inverse problem to determine the traction field
 97 at the cell-matrix interface, given the “experimentally measured” displace-
 98 ments. We also present the essential ingredients of our solution strategy
 99 which include a gradient-based minimization algorithm, an adjoint equation,
 100 and a special regularization approach. In Section 4, we describe three nu-
 101 merical examples that include tractions exerted on a hydrogel by a microglial
 102 cell and a neuronal cell. We use these examples to quantify and understand
 103 errors in the reconstructed traction field due to noise as well as incorrect
 104 model selection. We conclude with a summary in Section 5.

105 **2. The forward elasticity problem**

106 A typical problem of interest consists of a cell embedded within a three-
 107 dimensional scaffold made from a hydrogel. It is assumed that the mechan-
 108 ical properties of the cell are not known, while that of the gel are known
 109 [7, 8, 10, 23]. In this study, the hydrogel around the cell is modeled as an
 110 incompressible hyperelastic solid using a modified version of the Blatz (or

the Veronda-Westmann) model. Treating hydrogel as incompressible is common [7, 8, 10, 23]. In addition, several commonly used hydrogels, such as agar-gelatin co-gels, display significant nonlinear elastic behavior in range of strains encountered in cell-traction studies. For these types of gels the modified Blatz model is a simple constitutive model (with just 2 parameters) that captures the nonlinear stress-strain response.

2.1. Strong form

The problem is posed on the domain obtained by subtracting the cell from the hydrogel. Typically, the extent of the hydrogel is too large to fully model in this problem, so the domain is truncated at a certain distance away from the cell where the displacements (due to cellular tractions) have decayed significantly. This truncation is common [10, 20, 48], and the truncation distance usually depends on how fast the traction decays. Authors in [9, 10, 48] have presented some nice illustrations of the decay of tractions.

The displacement field, $\mathbf{u}$, and the pressure, p , within the hydrogel obey the equations of equilibrium given by:

$$\nabla \cdot \mathbf{P} = \mathbf{0} \quad \text{in } \Omega_0, \quad (1)$$

$$\mathbf{u} = \mathbf{g} \quad \text{on } \Gamma_g, \quad (2)$$

$$\mathbf{P} \cdot \mathbf{N} = \mathbf{h} \quad \text{on } \Gamma_h, \quad (3)$$

along with the incompressibility constraint:

$$J - 1 = 0, \quad \text{in } \Omega_0. \quad (4)$$

Here Eq.(1) is a statement of equilibrium in the reference configuration Ω_0 , where $\mathbf{P}$ is the first Piola-Kirchhoff stress tensor, and Eqs.(2) and (3) define the Dirichlet (displacement) and Neumann (traction) boundary conditions, respectively, where the displacement data, $\mathbf{g}$, is prescribed on Γ_g , and the traction data, $\mathbf{h}$, is prescribed on Γ_h . Further, $\partial\Omega_0 = \overline{\Gamma_g} \cup \overline{\Gamma_h}$ defines the closed boundary of Ω_0 , $\Gamma_g \cap \Gamma_h = \emptyset$, and $\mathbf{N}$ is the outward normal vector on the surface, Γ_g , in the reference configuration. Eq.(4) enforces the incompressibility constraint, and $J = \det(\mathbf{F})$ is the jacobian of the deformation gradient, which is defined as $\mathbf{F} = \nabla \mathbf{u} + \mathbf{1}$.

These equations are appended with a constitutive equation for the stress. In particular, first Piola-Kirchhoff stress tensor is given by $\mathbf{P} = \mathbf{F}\mathbf{S}$, where $\mathbf{S}$, the second Piola-Kirchhoff stress tensor, is defined as, $\mathbf{S} = -J \times p \mathbf{F}^{-1} \mathbf{F}^{-T} + 2 \frac{\partial W}{\partial \mathbf{C}}$. In the expression above, W is the strain energy density function that

141 depends on the invariants of the right Cauchy-Green strain tensor, $\mathbf{C} =$
 142 $\mathbf{F}^T \mathbf{F}$. For the constitutive model used in this paper, W is given by

$$W = \frac{\mu}{2\gamma} (e^{\gamma(J^{-\frac{2}{3}} I_1 - 3)} - 1), \quad (5)$$

143 where μ is the shear modulus at zero strain, and γ is the nonlinear parameter
 144 that determines the nonlinear change of stress with strain. Further, $I_1 =$
 145 $\text{trace}(\mathbf{C})$, is the first principal invariant of the Cauchy-Green strain tensor.
 146 More details about this material model can be found in [34, 49–51].

147 2.2. Weak form

148 We solve the forward problem using the finite element method, which is
 149 based on the weak, or the variational form, of the forward problem. The
 150 variational form may be derived by either minimizing the total potential
 151 energy of the system under the constraint of incompressibility, or it may
 152 be derived by multiplying the equilibrium equation and the incompress-
 153 ibility equations with weighting functions, integrating over Ω_0 , performing
 154 integration-by-parts on the former, and then imposing the traction bound-
 155 ary condition Eq.(3). In either case, we arrive at the following statement:
 156 Find $\mathbf{U} \equiv [\mathbf{u}, p] \in \mathcal{S} \times \mathcal{P}$ such that

$$\mathcal{A}(\mathbf{W}, \mathbf{U}) = (\mathbf{w}, \mathbf{h})_{\Gamma_h}, \quad \forall \mathbf{W} \equiv [\mathbf{w}, q] \in \mathcal{V} \times \mathcal{P}, \quad (6)$$

157 where

$$\mathcal{A}(\mathbf{W}, \mathbf{U}) \equiv \int_{\Omega_0} \nabla \mathbf{w} : \mathbf{P} \, d\Omega + \int_{\Omega_0} q(J - 1) \, d\Omega, \quad (7)$$

$$(\mathbf{w}, \mathbf{h})_{\Gamma_h} \equiv \int_{\Gamma_h} \mathbf{w} \cdot \mathbf{h} \, d\Gamma. \quad (8)$$

158 The function spaces $\mathcal{V}$, $\mathcal{S}$, and $\mathcal{P}$ that appear in the equations above are
 159 defined as,

$$\mathcal{V} = \{\mathbf{w} | w_i \in H^1(\Omega_0); w_i = 0 \text{ on } \Gamma_g\}, \quad (9)$$

$$\mathcal{S} = \{\mathbf{u} | u_i \in H^1(\Omega_0); u_i = g_i \text{ on } \Gamma_g\}, \quad (10)$$

$$\mathcal{P} \subseteq L_2(\Omega_0), \quad (11)$$

160 where L_2 is the space of square-integrable functions, and H^1 is the Sobolev
 161 space of square-integrable functions with square-integrable derivatives.

162 The standard Galerkin finite element method is obtained by restricting
 163 the weak form (Eq.6) to finite-dimensional function spaces. However, for
 164 incompressible materials, it is well known that the Galerkin finite element
 165 method leads to unstable solutions unless very specific choices are used for the
 166 displacement and pressure function spaces. This restriction can be overcome
 167 by stabilizing this formulation with residual-based terms [52, 53]. With that
 168 addition, the discrete stabilized formulation is given by: Find $\mathbf{U}^h \equiv [\mathbf{u}^h, p^h] \in$
 169 $\mathcal{S}^h \times \mathcal{P}^h$ such that

$$\mathcal{A}_s(\mathbf{W}^h, \mathbf{U}^h) = (\mathbf{w}^h, \mathbf{h})_{\Gamma_h}, \quad \forall \mathbf{W}^h \equiv [\mathbf{w}^h, q^h] \in \mathcal{V}^h \times \mathcal{P}^h, \quad (12)$$

170 where

$$\mathcal{A}_s(\mathbf{W}^h, \mathbf{U}^h) \equiv \mathcal{A}(\mathbf{W}^h, \mathbf{U}^h) - \sum_{e=1}^{n_{el}} (\tau \nabla \cdot (\mathbf{F}^h \mathbf{S}^h), \mathbf{F}^{h-T} \nabla q^h)_{\Omega_0^e}. \quad (13)$$

171 The second term in Eq.13 represents the stabilization term. The subscript
 172 Ω_0^e indicates that the L_2 inner product is evaluated in the interior of the e -
 173 th element, and the stabilization factor, $\tau = \frac{\alpha h^2}{2\mu}$, where $\alpha \approx 0.5$, and h is
 174 the characteristic element length. A detailed discussion of this stabilization
 175 method can be found in [34]. In our implementation, all variables, including
 176 displacement, pressure and traction, are represented by bilinear finite element
 177 basis functions.

178 3. Inverse problem for the traction vector field

179 In this section, we consider the inverse problem of determining the trac-
 180 tion field, $\mathbf{h}$, on the boundary, Γ_h , given measured displacements at discrete
 181 locations in the hydrogel. The overall scheme to obtain the solution of this
 182 inverse problem is similar to that of the material property identification in-
 183 verse problem described in [34] and [29]. The key difference is that we wish
 184 to recover the traction, $\mathbf{h}$, and not the material properties. The inverse
 185 problem is formulated as a minimization problem subject to the constraint
 186 of satisfying the forward problem. A gradient-based optimization approach,
 187 L-BFGS [54], is used to solve the minimization problem iteratively and the
 188 gradients are efficiently evaluated by solving the appropriate adjoint equa-
 189 tion [26]. Similar use of the adjoint method to efficiently compute gradients
 190 can also be found in other applications [55–58].

191 3.1. Inverse problem statement

192 The inverse problem is stated as follows: Given the measured displace-
193 ment field, $\tilde{\mathbf{u}}$, find the traction field, $\mathbf{h}$, such that the objective function

$$\pi(\mathbf{u}, \mathbf{h}) = \frac{1}{2} \|\mathbf{T}\mathbf{u} - \mathbf{T}\tilde{\mathbf{u}}\|_0^2 + \alpha \mathcal{R}(\mathbf{h}) \quad (14)$$

194 is minimized subject to the constraint that the predicted displacement field,
195 $\mathbf{u}$, satisfies Eqs.(1)-(4).

196 In Eq.(14), the first term is the displacement matching term. The mis-
197 match between the predicted and measured displacement field is measured
198 in the L_2 norm denoted by $\|\cdot\|_0$. It is weighted by the tensor $\mathbf{T}$, which can
199 be used to augment the contributions from the more accurate components
200 of measured displacements. The second term of Eq.(14) is the regularization
201 term, where α is the regularization parameter. The regularization term em-
202 beds prior information about the unknown field, in our case the traction field,
203 $\mathbf{h}$, into the inverse problem. In most cases, it is assumed that the recovered
204 field is smooth in a sense defined by the choice of this term. This has the
205 effect of regularizing the inverse problem, which is typically ill-posed, as it
206 lacks uniqueness or stability. In this manuscript, Tikhonov (H_1) regulariza-
207 tion [59] is implemented:

$$\mathcal{R}(\mathbf{h}) = \frac{1}{2} \int_{\Gamma_h} |\nabla_\Gamma \mathbf{h}|^2 d\Gamma. \quad (15)$$

208 In the equations above, ∇_Γ represents the gradient operator restricted to the
209 surface Γ_h . At any location $\mathbf{x} \in \Gamma_h$, it is given by $\nabla_\Gamma \equiv (\mathbf{1} - \mathbf{N} \otimes \mathbf{N})\nabla$,
210 where $\mathbf{1}$ is the identity tensor, $\mathbf{N}$ is the outward normal to the surface and
211 ∇ is the gradient operator. We note that ∇_Γ only retains the tangential
212 components of ∇ .

213 3.2. Formulation of the gradient

214 In order to solve the inverse problem using a quasi-Newton method, we
215 need to evaluate the derivative of π with respect to $\mathbf{h}$. This is relatively
216 easily evaluated for the regularization term. However, its evaluation for the
217 displacement mismatch term is cumbersome. This is because the predicted
218 displacement field, $\mathbf{u}$, depends upon $\mathbf{h}$ implicitly through the constraint of
219 Eq.(12). This difficulty is circumvented through the use of an appropriately

220 chosen adjoint field, which is derived by constructing the Lagrangian corre-
 221 sponding to π . That is,

$$L(\mathbf{U}^h, \mathbf{W}^h, \mathbf{h}) = \pi(\mathbf{u}^h, \mathbf{h}) + \mathcal{A}_s(\mathbf{W}^h, \mathbf{U}^h) - (\mathbf{w}^h, \mathbf{h})_{\Gamma_h}, \quad (16)$$

222 where $\mathbf{W}^h \in \mathcal{V}^h \times \mathcal{P}^h$ and $\mathbf{U}^h \in \mathcal{S}^h \times \mathcal{P}^h$. Here $\mathbf{W}^h$ plays the role of a
 223 Lagrange multiplier field.

224 We denote the variation of a function $f(x)$ in the direction δx by δf ,

$$\delta f = D_x f \cdot \delta x = \left. \frac{d}{d\epsilon} f(x + \epsilon \delta x) \right|_{\epsilon \rightarrow 0}. \quad (17)$$

225 The variation of L is given by

$$\delta L = D_{\mathbf{U}^h} L \cdot \delta \mathbf{U}^h + D_{\mathbf{W}^h} L \cdot \delta \mathbf{W}^h + D_{\mathbf{h}} L \cdot \delta \mathbf{h}. \quad (18)$$

226 Setting $D_{\mathbf{W}^h} L \cdot \delta \mathbf{W}^h = 0$, $\forall \delta \mathbf{W}^h \in \mathcal{V}^h \times \mathcal{P}^h$ in the above equation imme-
 227 diately yields an equation for $\mathbf{u}^h$, that is

$$\mathcal{A}_s(\delta \mathbf{W}^h, \mathbf{U}^h) = (\delta \mathbf{w}^h, \mathbf{h})_{\Gamma_h}, \quad \forall \delta \mathbf{W}^h \in \mathcal{V}^h \times \mathcal{P}^h. \quad (19)$$

228 This implies that $\mathbf{U}^h$ satisfies the forward problem Eq.(12). Under this con-
 229 dition, from Eq.(16), we conclude $\pi = L$, and hence $\delta \pi = \delta L$.

230 We now examine the condition for which the expression for δL can be
 231 further simplified. In particular, we set $D_{\mathbf{U}^h} L \cdot \delta \mathbf{U}^h = 0$, $\forall \delta \mathbf{U}^h \in \mathcal{V}^h \times \mathcal{P}^h$
 232 to arrive at

$$\mathcal{B}_s(\mathbf{W}^h, \delta \mathbf{U}^h; \mathbf{U}^h) = -(\mathbf{T} \delta \mathbf{u}, \mathbf{T} \mathbf{u} - \mathbf{T} \tilde{\mathbf{u}}), \quad \forall \delta \mathbf{U}^h \in \mathcal{V}^h \times \mathcal{P}^h. \quad (20)$$

233 where $\mathcal{B}_s(\mathbf{W}^h, \delta \mathbf{U}^h; \mathbf{U}^h) \equiv \left. \frac{d}{d\epsilon} \mathcal{A}_s(\mathbf{W}^h, \mathbf{U}^h + \epsilon \delta \mathbf{U}^h) \right|_{\epsilon \rightarrow 0}$ is the linearization of
 234 $\mathcal{A}_s(\cdot, \cdot)$ about $\mathbf{U}^h$. Eq.(20) yields a well-posed linear problem for $\mathbf{W}^h$. The
 235 left hand side of this problem is the adjoint of the linearized version of the
 236 original forward problem Eq.(12). For this reason, this problem is referred
 237 to as the adjoint problem, and $\mathbf{W}^h$ the adjoint field.

238 With $\mathbf{U}^h$ given by the primal problem Eq.(19) and $\mathbf{W}^h$ given by the
 239 adjoint problem Eq.(20), we have from Eq.(18)

$$\delta L = \delta \pi = D_{\mathbf{h}} L \cdot \delta \mathbf{h} = \alpha D_{\mathbf{h}} \mathcal{R} \cdot \delta \mathbf{h} - (\mathbf{w}^h, \delta \mathbf{h})_{\Gamma_h}, \quad (21)$$

240 where $\mathcal{R}$ is the regularization term defined by Eq.(15). As a result, the
 241 algorithm to calculate the change in the objective function corresponding to
 242 a change in the traction vector proceeds as follows:

- 243 1. Solve Eq.(19) to determine the primal field $\mathbf{U}^h = [\mathbf{u}^h, p^h]$.
- 244 2. Solve Eq.(20) to determine the adjoint field $\mathbf{W}^h = [\mathbf{w}^h, q^h]$.
- 245 3. Use $\mathbf{w}^h$ in Eq.(21) to determine the variation in the objective function.

246 In this study, the optimization variables are the traction components at
 247 each node on the cell-matrix interface. The value of the objective function,
 248 Eq.(14), and its gradient, Eq.(21), are the input for the L-BFGS optimization
 249 algorithm, which returns an updated estimate of tractions for each node and
 250 the whole process is repeated until convergence or until the maximum number
 251 of iterations is achieved.

252 *Remark*

253 We note that the primal problem, Eq.(19), is a system of nonlinear equa-
 254 tions which is solved using the Newton method. This represents the major
 255 computational cost of our algorithm. This cost is kept in check by utilizing a
 256 continuation scheme described in [34] and [29]. Within this approach, when
 257 solving the forward traction problem for a new guess of the traction field,
 258 the solution from the previous step is used as an initial state. Consequently,
 259 the forward problem is solved within 10 Newton iterations, as opposed to on
 260 the order of 100 iterations otherwise.

261 4. Numerical examples

262 In this section, two geometrically accurate cell models are used to demon-
 263 strate the applicability of our nonlinear algorithm in recovering cell traction.
 264 In each case, a known traction field, $\hat{\mathbf{h}}$, is applied at the cell-matrix interface
 265 and the forward problem is solved. This results in the displacement of the
 266 matrix which is sampled at some select nodes. We denote this displacement
 267 vector by $\hat{\mathbf{u}}^{(j)}, j = 1, \dots, n_{node}$. In some cases, in order to simulate the effect
 268 of instrument noise and errors in the displacement estimation algorithm in
 269 actual experiments, noise is added to this displacement. The noisy “mea-
 270 sured” displacement field is given by

$$\tilde{\mathbf{u}}^{(i)} = \hat{\mathbf{u}}^{(i)} + \eta \times \frac{\sum_{j=1}^{n_{node}} |\hat{\mathbf{u}}^{(j)}|}{\sum_{j=1}^{n_{node}} |\mathbf{n}^{(j)}|} \times \mathbf{n}^{(i)}, i = 1, \dots, n_{node} \quad (22)$$

271 where $|\cdot|$ denotes the magnitude of a vector, $n_k^{(j)} \in N(0, 1)$, $k = 1, 2, 3$
 272 and $N(0, 1)$ is standard normal distribution. In this case, η , represents the

fraction of noise in the “measured” displacement field. The effect of this parameter on the quality of the reconstruction is examined.

In order to quantify the performance of our algorithm, the difference between the recovered traction field, $\mathbf{h}$, and the applied traction field, $\hat{\mathbf{h}}$, is measured,

$$e(\mathbf{h}) = \frac{\|\mathbf{h} - \hat{\mathbf{h}}\|_0}{\|\hat{\mathbf{h}}\|_0}. \quad (23)$$

For simplicity, a weight of unity is selected for the entire displacement vector, which means that the tensor, $\mathbf{T}$, in Eq.(14) is set to the identity tensor. In order to be unbiased, the initial guess for the traction field is set to 0. Also, the regularization parameter for the following results in this paper was obtained by solving the inverse problem with different values of the regularization parameter, and plotting the variation of displacement mismatch term as a function of the regularization parameter (the so-called L-curve [60]). See Figure 4, for the L-curve for a typical case considered in this study. The quasi-Newton iterations were considered “converged” when the relative change in the displacement matching term over the last five iterations was smaller than 1×10^{-8} .

4.1. Microglial cell model

The 3D model of the microglial cell (see Figure 1a) is reconstructed through a set of pictures of z-stack microscopy (image courtesy of Sebastian Rhode at Carl Zeiss Microscopy and Sabine Scheibe) with ImageJ [61] and meshed with Simmodeler (Simmertrix.Inc, New York, US). The cell is about $20 \mu\text{m} \times 20 \mu\text{m} \times 15 \mu\text{m}$ and is contained within a cubical matrix with an edge length of $100 \mu\text{m}$. The finite element mesh is comprised of 9,641 vertices and 51,544 tetrahedral elements. Out of these, the vertices that are on the surface of the cell, and beyond a sphere of $50 \mu\text{m}$ diameter, are treated as location without beads. That is the displacement field from these locations is not used in solving the inverse problem. This gives us a total of 7,023 bead locations to work with. The exclusion of other locations helps us in avoiding the “inverse crime” of using all the forward data in the inverse problem. It is also consistent with the experimental protocol where most of the beads that are close to the cell are tracked [62]. We note that the number of beads considered in this study is comparable to experiments [10] and is much more than some of the state-of-the-art cell traction recovery algorithms can handle [12, 20].

307 In the numerical experiments reported here, the shear modulus, μ , of
 308 the matrix is set to either 400 Pa or 600 Pa. The values lie within the
 309 range of commonly used experimental hydrogels, extracellular matrices and
 310 real tissues [10, 63, 64]. The dimensionless nonlinear parameter, γ , in the
 311 modified blatz model is set to either 1 or 5. With $\gamma = 1$ the material behaves
 312 linearly up to about 100% strain, and with $\gamma = 5$ it starts to deviate from
 313 linear behavior at about 15% strain [34]. All six surfaces of the box are
 314 assumed fixed in the x , y and z directions ($u_x = u_y = u_z = 0$).

315 As shown in Figure 1b, the traction field is applied such that it is max-
 316 imum along the two protrusions of the cell, and is smoothly interpolated
 317 between these regions. The direction of the traction field is roughly pointed
 318 to the center of the cell. This loading condition simulates the contraction of
 319 the matrix close to the cell after the cell is lysed with detergent [10].

320 In the following sections, we consider the effect of material and geometric
 321 nonlinearity in recovering the traction vector field. In the first section, we
 322 set $\gamma = 1$ so that within the range of strains considered, the matrix behaves
 323 like a linear elastic solid. Thereafter, we consider $\gamma = 5$, where the material
 324 displays a nonlinear elastic response. We note that while gelatin substrates
 325 tend to be linear elastic for large values of strain, gelatin-agar substrates
 326 display nonlinear elastic behavior at relatively small ($\approx 15\%$) strains [18].

327 *Linear material ($\gamma = 1$)*

328 The shear modulus for the matrix is set to 400 Pa. In Figure 1b, we
 329 observe that the largest tractions are applied at the tips of the cell, and the
 330 magnitude drops as we approach the center of the cell. The traction pulls
 331 each arm of the cell toward the center of the cell. The resulting displacement
 332 fields are maximum at the tip of the arms, and at the connections between
 333 arms and the rest of the cell. Significant strains are generated on the cell-
 334 matrix interface, and are illustrated in Figures 1c, where we have plotted the
 335 first principal component of the Green-Lagrange strain tensor. We note that
 336 this strain ranges from about 50% to 90% on the arms - Figure 1d. The large
 337 strain indicates that the use of a linear measure of strain is inappropriate for
 338 this problem.

339 For each set of “measured” displacements (with 1% and without noise),
 340 the traction field was reconstructed using either a finite strain or an infinites-
 341 imal strain model. The resulting traction vector fields are shown in Figure
 342 2. We observe that while the spatial variation of these fields is very similar,
 343 their magnitudes are significantly different. This is made clear by computing

the error, $e(\mathbf{h})$, in the reconstructed traction field (see Figure 3). For the no-noise case with the finite strain model, this error is 4.23%, and with the infinitesimal strain model it is 12.7% (regularization parameters = 0.0001 and 1, respectively). When 1% noise is added to the displacements, the error is 12.75% for the finite strain model, and 20.7% for the infinitesimal strain model (regularization parameters = 5 and 10, respectively). The difference between these two errors, which is around 8%, is therefore the error incurred in incorrectly making the infinitesimal strain assumption. Here we note that the infinitesimal strain assumption is often made [7–10, 12, 20]. Figure 4 displays the L-curve for the case with finite deformation and 1% noise as a typical curve for the cases considered in this paper. According to [60], the value of the regularization parameters at the point of largest curvature is the “optimal” regularization parameter. For the example considered in this curve, this corresponds to $\alpha = 5$ (circled in blue in Figure 4). We use this approach to find the regularization parameters for all cases in this paper. For the sake of brevity, we do not show all the L-curve plots.

Nonlinear material ($\gamma = 5$)

In this case when solving the forward problem, in order to generate the “measured” data, the shear modulus for the matrix is set to 600 Pa, and the nonlinear parameter is set to $\gamma = 5$. The traction field is identical to the $\gamma = 1$ case described above. The traction field and the displacement magnitude are shown in Figure 5a. We observe that when compared with the linear elastic case, the displacement for this case is smaller. In examining the image for the principal component of the Green-Lagrange strain, we observe that the maximum strain is also much smaller (approximately 40% as opposed to 90%). This is attributed to the higher shear modulus and the larger values of the nonlinear parameter in this case. This leads to a large value of the tangent modulus for the matrix, particularly at large values of strain. Given that the traction vector field for the two cases is the same, a larger elastic tangent modulus implies a small overall strain.

Once again two cases were considered: one with no noise and one with 1% noise. For each case, two inverse problems were solved. In one problem, the matrix deformation was treated with a finite strain theory and the appropriate nonlinear elastic model was utilized. In the second problem, the deformation was incorrectly treated as linear elastic, and an infinitesimal strain model was used. The resulting traction vectors are shown in Figure 6. The spatial distribution of these fields is very close to the “exact” trac-

tion field, however their magnitudes are different. Their departures from the applied traction field are quantified in Figure 3. In the no-noise case, the errors, when using the finite strain and infinitesimal strain models, are 4.5% and 14.76%, respectively (regularization parameter = 0.0001 and 1, respectively). The difference between the two is significant. For the 1% noise case, the corresponding errors are 18.66% and 27.23% (regularization parameter = 5 and 10, respectively), and the difference is around 9%. Once again we observe that the model error, that is the error associated with assuming an incorrect model in the inverse problem, is significant.

4.2. Neuronal cell model

As the second example, the geometry of a multi-polar neuronal cell is considered (see Figure 7a). Recently, there has been significant interest in understanding the traction exerted by these cells [65, 66]. The size of the neuronal cell is about $10\ \mu\text{m} \times 10\ \mu\text{m} \times 10\ \mu\text{m}$, and it is contained within a cube of length $80\ \mu\text{m}$. The unstructured mesh for this configuration consists of 54,587 elements with 10,673 vertices. Out of these, 6,971 are treated as tracked beads (that is the displacement at these locations are “measured”). These locations are selected so that they are close to the cell-matrix interface (but not on it). The shear modulus of the surrounding matrix is 400 Pa, and the nonlinear parameter is set to either 1 or 5. Five of the outer surfaces of the box are fixed in the x , y and z directions ($u_x = u_y = u_z = 0$), and the sixth surface is traction free, which is to mimic the free top surface of the matrix in experiments.

As shown in Figure 7b, the traction field is selected such that its magnitude is largest along the arms of the cell and smoothly reduces towards the center. In addition, the traction vector is pointed inward toward the center of the cell. Similar to the first example, this loading condition also simulates the contraction of the matrix close to the cell after the cell is lysed with detergent [10].

Linear material ($\gamma = 1$)

In Figure 7c, we observe that the largest strain occurs at the dendritic ends of the cell, and pulls the dendrites toward the center of the cell body. Significant strains (50% to 90% along the dendrites) are generated at the cell-matrix surface, and are illustrated in Figure 7d, where we have plotted the first principal component of the Green-Lagrange strain tensor.

416 When solving the inverse problem, we use displacements with no noise and
 417 1% noise, and then either use a finite strain or an infinitesimal strain model.
 418 The recovered traction vector fields appear similar, and are shown in Figure
 419 8. However, their magnitudes are different. In Figure 9, we quantify this by
 420 evaluating the error ($e(\mathbf{h})$) in the traction field for each case. For “measured”
 421 displacement with no noise, this error is 0.5% for the finite strain model, and
 422 16.93% for the infinitesimal strain model (regularization parameters = $1 \times$
 423 10^{-7} and 0.1, respectively). For “measured” displacement with 1% noise, the
 424 error for the finite strain model grows to 26.41%, and that for the infinitesimal
 425 strain model is 34.78% (regularization parameters = 0.5 and 1, respectively).
 426 The difference between these errors is the penalty associated with invoking
 427 the infinitesimal strain assumption.

428 *Nonlinear material ($\gamma = 5$)*

429 The problem described above is repeated with the nonlinear parameter
 430 now set to 5. This means that the matrix now stiffens with increasing strain,
 431 and given that the traction field is unchanged, it strains less in response to
 432 the traction exerted by the cell. This can be seen by examining Figures 10a
 433 and 10b where we have plotted the displacement magnitude and the principal
 434 component of the Green-Lagrange strain tensor on the cell-matrix interface.

435 Once again displacement fields with no noise and with 1% noise are con-
 436 sidered, and when solving the inverse problem, a finite strain model (with
 437 the appropriate nonlinear elastic response) and an infinitesimal strain model
 438 (with a linear elastic response) is used. This is done to quantify the effect
 439 of noise and modeling assumptions in solving the inverse problem. In all
 440 cases, we observe that the spatial distributions of the traction vector is re-
 441 covered with remarkable accuracy (see Figure 11, regularization parameter
 442 = 1×10^{-7} , 0.5, 0.1 and 1, respectively). However, the magnitude of the
 443 traction, and consequently the error in the reconstruction (see Figure 9), is
 444 sensitive to both displacement noise, and model selection. When there is no
 445 noise, the contribution of the model error is greater than 20%, and with 1%
 446 noise it drops to around 8%.

447 *4.3. Discussion*

448 We have selected the microglial and neuron cells as examples because of
 449 their biomedical relevance, and also because of their distinct shapes. While
 450 the microglial cell has a relatively smooth shape with very few protrusions,
 451 the shape of the neuron is much more complicated with several dendritic

452 structures. We note that in both cases our algorithm could be applied suc-
 453 cessfully. However, for the no-noise cases the tractions for the neuron cell
 454 were recovered with almost no error, whereas for the microglial cell they were
 455 recovered to around 4% error. There are several possible explanations for this
 456 difference. First, the magnitude of tractions applied to the microglial cell is
 457 about two times that of the tractions applied to the neuron, and in both
 458 cases the inverse problem begins with a zero initial guess. Consequently,
 459 for the microglial cell quasi-Newton algorithm has to cover a larger range
 460 in order to achieve the exact value. Also the dendritic shape of the neuron
 461 implies that there are many more local regions where the strains are large
 462 and these regions are sampled by the tracking beads. This could imply that
 463 the measured displacement field for the neuron carries a “stronger” imprint
 464 of the traction field making the inverse problem better posed.

465 In biomechanics applications, often the tractions measured are used to
 466 determine average quantities such as the contractile moment of the cell, or
 467 the net strain energy in the gel. We may conjecture how the proposed method
 468 would perform in evaluating these quantities. We note that the contractile
 469 moment is a linear functional of the traction field. Thus we would expect
 470 that error observed in this quantity would be similar to the error observed
 471 in the traction field. On the other hand, the strain energy in the gel may
 472 be evaluated by computing the work done by the cell on the gel. That is
 473 the integral $\int_{\Gamma} \mathbf{h} \cdot \mathbf{u} d\Gamma$. In this integral, $\mathbf{h}$ is the reconstructed traction field,
 474 and $\mathbf{u}$ is the estimated displacement on the interface. It is reasonable to
 475 expect that if the recovered traction is underestimated by a certain fraction,
 476 then the recovered displacement will also be underestimated by the same
 477 factor. Then roughly speaking, the integral, which involves the product of
 478 these quantities, will be underestimated by two times this factor. In this
 479 case the importance of including nonlinear elastic effects will be even more
 480 significant.

481 5. Summary

482 Traction force microscopy is an important technique that is used to quan-
 483 tify tractions at the cell-matrix interface. It works by measuring the motion
 484 of embedded micro-beads in response to cellular tractions and using this
 485 information to infer the traction field. In this manuscript we have posed
 486 TFM as an inverse problem, and solved it using a constrained minimiza-
 487 tion algorithm. In doing so we have accounted for the three-dimensional

488 nature of the problem, the complex geometry of the cell, and the finite-
489 strain and nonlinear elastic behavior of the matrix. Through the use of a
490 quasi-Newton algorithm, a carefully derived adjoint problem, and a novel
491 continuation strategy, we have developed and implemented algorithms that
492 efficiently utilize data from a large number of tracking beads ($\approx 10^4$) and
493 reconstruct traction vectors on a well-resolved surface mesh ($\approx 10^3 - 10^4$
494 grid points).

495 We have applied these algorithms to *in-silico* problems with realistic ge-
496 ometric models of microglial and neuronal cells. We conclude that the pro-
497 posed algorithms are able to accurately recover the traction fields. We have
498 also tested the effect of measurement noise on the proposed methods, and
499 examined the error incurred by not including nonlinear effects when solving
500 the TFM problem. We conclude that the error in the recovered traction
501 field is sensitive to noise in the measured displacements, and to neglecting
502 nonlinear effects in the TFM problem.

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7. References

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Figure 1: Microglial cell: (a) Mesh on the exterior surface and the cell-matrix interface. Results of the simulated forward problem on the deformed surface with $\gamma = 1$. (b) Arrows display the traction field (in kPa), and the deformed surface is colored by the displacement magnitude in μm . (c) The first principal component of the Green-Lagrange strain tensor. (d) Distribution of the strain at the bead locations as a function of distance from the center.

Figure 2: Results of the inverse analysis for the microglial cell on the deformed surface with $\gamma = 1$. Arrows display the recovered traction field (in kPa), and the deformed surface is colored by the recovered displacement magnitude in μm . (a) Solution with nonlinear effects and 0% noise. (b) Solution with nonlinear effects and 1% noise. (c) Solution without nonlinear effects and 0% noise. (d) Solution without nonlinear effects and 1% noise.

Figure 3: Error summary for the microglial cell model.

Figure 4: L-curve for the microglial cell model with nonlinear effects and 1% noise.

(a)

(b)

(c)

Figure 5: Results of the simulated forward problem for the microglial cell model on the deformed surface with $\gamma = 5$. (a) Arrows display the traction field (in kPa), and the deformed surface is colored by the displacement magnitude in μm . (b) The first principal component of the Green-Lagrange strain tensor. (c) Distribution of the strain at the bead locations as a function of distance from the center.

Figure 6: Results of the inverse analysis for the microglial cell on the deformed surface with $\gamma = 5$. Arrows display the recovered traction field (in kPa), and the deformed surface is colored by the recovered displacement magnitude in μm . (a) Solution with nonlinear effects and 0% noise. (b) Solution with nonlinear effects and 1% noise. (c) Solution without nonlinear effects and 0% noise. (d) Solution without nonlinear effects and 1% noise.

Figure 7: Neuronal cell: (a) Mesh on the exterior surface and the cell-matrix interface. Results of the simulated forward problem on the deformed surface with $\gamma = 1$. (b) Arrows display the traction field (in kPa), and the deformed surface is colored by the displacement magnitude in μm . (c) The first principal component of the Green-Lagrange strain tensor. (d) Distribution of the strain at the bead locations as a function of distance from the center.

Figure 8: Results of the inverse analysis for the neuronal cell on the deformed surface with $\gamma = 1$. Arrows display the recovered traction field (in kPa), and the deformed surface is colored by the recovered displacement magnitude in μm . (a) Solution with nonlinear effects and 0% noise. (b) Solution with nonlinear effects and 1% noise. (c) Solution without nonlinear effects and 0% noise. (d) Solution without nonlinear effects and 1% noise.

Figure 9: Error summary for the neuron cell model.

(a)

(b)

(c)

Figure 10: Results of the simulated forward problem for the neuronal cell model on the deformed surface with $\gamma = 5$. (a) Arrows display the traction field (in kPa), and the deformed surface is colored by the displacement magnitude in μm . (b) The first principal component of the Green-Lagrange strain tensor. (c) Distribution of the strain at the bead locations as a function of distance from the center.

Figure 11: Results of the inverse analysis for the neuronal cell on the deformed surface with $\gamma = 5$. Arrows display the recovered traction field (in kPa), and the deformed surface is colored by the recovered displacement magnitude in μm . (a) Solution with nonlinear effects and 0% noise. (b) Solution with nonlinear effects and 1% noise. (c) Solution without nonlinear effects and 0% noise. (d) Solution without nonlinear effects and 1% noise.